



(86) Date de dépôt PCT/PCT Filing Date: 2001/06/05
(87) Date publication PCT/PCT Publication Date: 2002/01/24
(45) Date de délivrance/Issue Date: 2009/08/25
(85) Entrée phase nationale/National Entry: 2002/03/18
(86) N° demande PCT/PCT Application No.: JP 2001/004731
(87) N° publication PCT/PCT Publication No.: 2002/006482
(30) Priorité/Priority: 2000/07/19 (JP2000-219652)

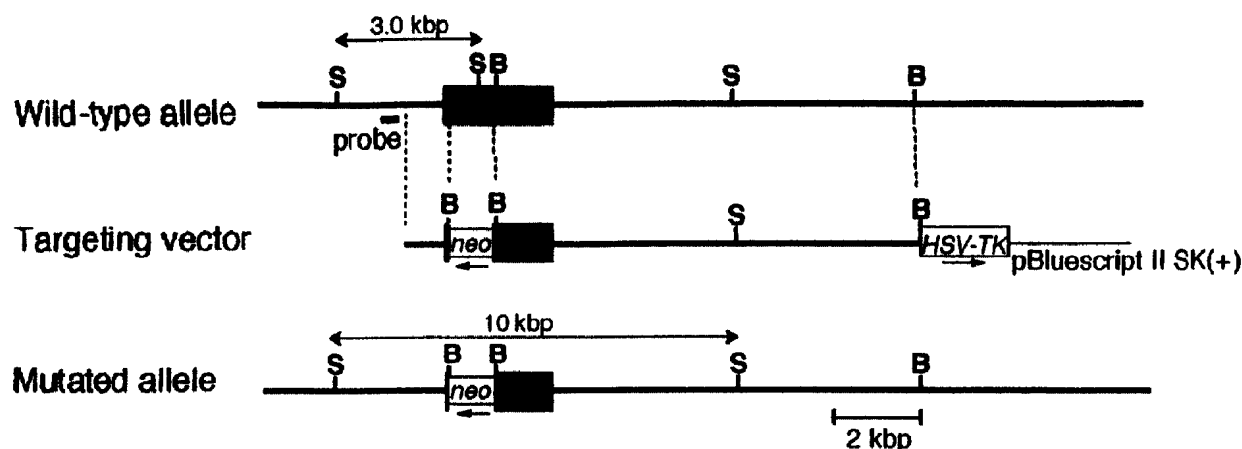
(51) Cl.Int./Int.Cl. *C12N 15/12* (2006.01),
A61K 38/00 (2006.01), *A61K 45/00* (2006.01),
A61P 31/04 (2006.01), *A61P 35/00* (2006.01),
A61P 37/08 (2006.01), *C07K 14/705* (2006.01),
C07K 16/28 (2006.01), *C12N 5/10* (2006.01),
C12P 21/02 (2006.01), *C12P 21/08* (2006.01),
C12Q 1/68 (2006.01), *G01N 33/15* (2006.01),
G01N 33/50 (2006.01), *G01N 33/566* (2006.01),
G01N 33/577 (2006.01)

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(54) Titre : PROTEINE RECEPTRICE RECONNAISSANT SPECIFIQUEMENT UN ADN BACTERIEN

(54) Title: RECEPTOR PROTEINS SPECIFICALLY RECOGNIZING BACTERIAL DNA



(57) **Abrégé/Abstract:**

The present invention provides a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence, a genomic DNA encoding it, an experimental animal model useful for examining responsiveness of a host immune cell against a bacterial infectious disease. DNA encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is screened by BLAST search, a number of EST clones having high homology with various TLRs is screened, these clones are used as a probe to isolate a full-length cDNA from mouse macrophage cDNA library, and the sequence of bases of the cDNA is analyzed to confirm that it is TLR9 comprising a conserved regions such as LRR and TIR regions, and then a knockout mouse is produced to confirm that TLR9 is a receptor protein of oligonucleotides having an unmethylated CpG sequence of bacterial DNA.

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ABSTRACT

The present invention provides a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence, a genomic DNA encoding it, an experimental animal model useful for examining responsiveness of a host immune cell against a bacterial infectious disease. DNA encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is screened by BLAST search, a number of EST clones having high homology with various TLRs is screened, these clones are used as a probe to isolate a full-length cDNA from mouse macrophage cDNA library, and the sequence of bases of the cDNA is analyzed to confirm that it is TLR9 comprising a conserved regions such as LRR and TIR regions, and then a knockout mouse is produced to confirm that TLR9 is a receptor protein of oligonucleotides having an unmethylated CpG sequence of bacterial DNA.

SPECIFICATION

RECEPTOR PROTEINS SPECIFICALLY RECOGNIZING BACTERIAL DNA

TECHNICAL FIELD

The present invention relates to a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence, a gene of the receptor protein and uses of them.

BACKGROUND OF THE INVENTION

It is already known that Toll genes are necessary for determining the dorsoventral axis in the embryogeny of *Drosophila* (Cell 52, 269-279, 1988, Annu. Rev. Cell Dev. Biol. 12, 393-416, 1996) and for antifungal immune responses in the adult fly (Cell 86, 973-983, 1996).

It has been shown that the Toll is a Type I transmembrane receptor comprising leucine-rich repeat (LRR) in extracellular domains, and its intracellular domains are highly homologous to the intracellular domains of mammalian interleukin-1 receptor (IL-1R) (Nature 351, 355-356, 1991, Annu. Rev. Cell Dev. Biol. 12, 393-416, 1996, J. Leukoc. Biol. 63, 650-657, 1998).

Recently, mammalian homologs of Toll called Toll-like Receptor (TLR) have been identified, and six members of the family such as TLR2 and TLR4 have been reported (Nature 388, 394-397, 1997, Proc. Natl. Acad. Sci. USA 95, 588-593, 1998, Blood 91, 4020-4027, 1998, Gene 231, 59-65, 1999). It is known that a member of the TLR family mediates MyD88, an adapter protein as IL-1R is, recruits IL-1R binding kinase (IRAK), activates TRAF6, and activates downstream NF- κ B (J. Exp. Med. 187, 2097-2101, 1998, Mol. Cell 2, 253-258, 1998, Immunity 11, 115-122, 1999). It is also thought that the role of the TLR family in mammals is related to innate immune recognition as a pattern recognition receptor (PRR) recognizing bacterial common components (Cell 91, 295-298, 1997).

It is well known that one of the pathogen-associated molecular patterns (PAMP) recognized by the PRR mentioned above is lipopolysaccharide (LPS), which is a main component of the outer membrane of Gram-negative bacteria (Cell 91, 295-298, 1997), the LPS stimulates a host cell to produce various inflammatory cytokines such as TNF α , IL-1 or IL-6 in the host cell (Adv. Immunol. 28, 293-450, 1979, Annu. Rev. Immunol. 13, 437-457,

1995), and the LPS captured by LPS-binding protein (LBP) is transferred to CD 14 on the surface of a cell (Science 249, 1431-1433, 1990, Annu. Rev. Immunol. 13, 437-457, 1995). The present inventors generated TLR4 knockout mice and reported that the TLR4 knockout mice lack the ability to respond to LPS, a main component of the outer membrane of the Gram-negative bacteria (J. Immunol. 162, 3749-3752, 1999), and also generated TLR2 knockout mice and reported that macrophages derived from TLR2 knockout mice showed low levels of response to cell wall of Gram-negative bacteria or peptidoglycan, a component of the Gram-negative bacteria (Immunity 11, 443-451, 1999).

On the other hand, from the fact that the oligonucleotides comprising bacterial DNA (DNA derived from bacteria) or an unmethylated CpG sequence stimulate immune cells of mice or human (Trends Microbiol. 4, 73-76, 1996, Trends Microbiol. 6, 496-500, 1998), and stimulate a T helper 1 cells (Th1)-like inflammatory response dominated by the release of IL-12 and IFN γ (EMBO J. 18, 6973-6982, 1999, J. Immunol. 161, 3042-3049, 1998, Proc. Natl. Acad. Sci. USA 96, 9305-9310, 1999), it is advocated that the oligonucleotides comprising CpG sequence are possibly used as an adjuvant in vaccine strategies including vaccines to cancer, allergy and infectious diseases (Adv. Immunol. 73, 329-368, 1999, Curr. Opin. Immunol. 12, 35-43, 2000, Immunity 11, 123-129, 1999). Although its effects have been expected in the clinical practice in this way, the molecular mechanism by which bacterial DNA comprising an unmethylated CpG sequence activates immune cells is unclear.

Although the DNA derived from bacteria comprising an unmethylated CpG motif activates immune cells significantly and induces response by Th1 as mentioned above, the activities at the molecular level are not well understood. The goal of the present invention is to provide a receptor protein TLR9, a member of TLR family specifically recognizing bacterial DNA comprising an unmethylated CpG sequence, the DNA encoding it, and the artificial animal models useful in examining response of host immune cells to bacterial infectious diseases, which elucidate effects of oligonucleotides comprising an unmethylated CpG sequence of bacterial DNA at the molecular level.

As a member of the mammalian TLR family, a pattern recognition receptor recognizing common structures of bacteria, relevant to innate immune recognition, six members (TLR1 to 6) have been publicized until now (Nature 388, 384-397, 1997, Proc. Natl. Acad. Sci. USA, 95, 588-593, 1998, Gene 231, 59-65, 1999), and TLR7 and TLR8, two novel members, are registered in GenBank (Registration No: AF240467 and AF246971). Although full-length

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cDNA is also found out for TLR9, and is registered in GenBank (Registration No: AF245704), its function has not been known.

The present inventors screened the DNA encoding
5 TLR family member receptor proteins specifically recognizing
bacterial DNA comprising an unmethylated CpG sequence on
BLAST search, screened a number of sequence tagged (EST)
clones highly homologous to various TLR already identified,
isolated full-length cDNA from mouse macrophage cDNA library
10 by using the fragments as a probe. The inventors also
isolated the human cDNA in the same manner. Next, the
sequences of bases of the cDNA were examined, and it was
confirmed that it is TLR9, in which regions conserved in the
TLR family such as LRR and TIR domains are present. We
15 generated TLR9 knockout mice, showed that TLR9 is a receptor
protein to the oligonucleotides comprising an unmethylated
CpG sequence of bacterial DNA and completed the invention.

DISCLOSURE OF THE INVENTION

A first aspect of the present invention relates to
20 DNA encoding a receptor protein specifically recognizing
bacterial DNA having an unmethylated CpG sequence. The
receptor protein specifically recognizing bacterial DNA
having an unmethylated CpG sequence may be (a) a protein
comprising the amino acid sequence shown in SEQ ID No: 2, or
25 (b) a protein comprising an amino acid sequence wherein one
or more of amino acids are deleted, substituted, or added in
the amino acid sequence shown in SEQ ID No: 2, and having
reactivity against bacterial DNA having an unmethylated CpG
sequence. The DNA preferably comprises the base sequence
30 shown in SEQ ID No: 1 or its complementary sequence, or part
or whole of the sequences. In one embodiment, the DNA

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hybridizes with the DNA comprising the base sequence shown in SEQ ID No: 1 under a stringent condition. In a certain embodiment, the DNA encodes (a) a protein comprising the amino acid sequence shown in SEQ ID No: 4, or (b) a protein
5 comprising an amino acid sequence wherein one or more of amino acid are deleted, substituted, or added in the amino acid sequence shown in SEQ ID No: 4, and having reactivity against bacterial DNA having an unmethylated CpG sequence. In one embodiment, the DNA comprises the base sequence shown
10 in SEQ ID No: 3 or its complementary sequence, or part or whole of the sequences. In one embodiment, the DNA hybridizes with the DNA comprising the base sequence shown in SEQ ID No. 3 or its complementary sequence under a stringent condition.

15 A second aspect of the present invention relates to a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence. The protein preferably comprises the amino acid sequence shown in SEQ ID No: 2 or an amino acid sequence wherein one or more of amino acids
20 are deleted, substituted or added in the amino acid sequence shown in SEQ ID No: 2. The protein may comprise the amino acid sequence shown in SEQ ID No: 4 or an amino acid sequence wherein one or more of amino acids are deleted, substituted or added in the amino acid sequence shown in SEQ
25 ID No: 4.

A third aspect of the present invention relates to a fusion protein comprising the protein of the second aspect fused with a marker protein and/or a peptide tag; an antibody, preferably a monoclonal antibody, which
30 specifically binds to the protein of the second aspect; and a host cell comprising an expression system expressing the protein of the second aspect.

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A fourth aspect of the present invention relates to a non-human animal. In a first embodiment, in the non-human animal, a gene encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is excessively expressed. In a second embodiment, in the non-human animal, a gene function encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is destroyed on a chromosome. Preferably, the non-human animal of the second embodiment has no reactivity against bacterial DNA having an unmethylated CpG sequence. The non-human animal is preferably a rodent animal, more preferably a mouse.

A fifth aspect the present invention relates to (1) a method of preparing a cell expressing a protein having reactivity against bacterial DNA having an unmethylated CpG sequence, in which the DNA of the first aspect is introduced into a cell wherein a gene function encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is destroyed on a chromosome; and (2) a cell expressing a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence obtained by the method.

A sixth aspect of the present invention relates to a screening method for an agonist or an antagonist of a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence. In a first embodiment, the method comprises: in vitro culturing a cell expressing a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence in the presence of a target substance, and measuring/evaluating TLR9 activity. In a second embodiment, the method comprises: administrating

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a target substance to a non-human animal wherein a gene function encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is destroyed on a chromosome, and

5 measuring/evaluating TLR9 activity of macrophages or spleen cells obtained from the non-human animal. In a third embodiment, the method comprises: administrating a target substance to a non-human animal wherein a gene encoding a receptor protein specifically recognizing bacterial DNA

10 having an unmethylated CpG sequence is excessively expressed, and measuring/evaluating TLR9 activity of macrophages or spleen cells obtained from the non-human animal. The non-human animal is preferably a mouse.

A seventh aspect of the present invention relates

15 to an agonist or an antagonist of a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence obtained by the screening method of the sixth aspect; a pharmaceutical composition comprising whole or part of a receptor protein specifically recognizing

20 bacterial DNA having an unmethylated CpG sequence as an active component: a pharmaceutical composition comprising the agonist or antagonist as an active component; a kit used to diagnose diseases related to the deletion, substitution and/or addition in a sequence of DNA encoding a receptor

25 protein specifically recognizing bacterial DNA having an unmethylated CpG sequence comprising a specific DNA comprising the base sequence shown in SEQ ID No: 1 or its complementary sequence or a part of one of these sequences, which can compare a sequence of DNA encoding a receptor

30 protein specifically recognizing bacterial DNA having an unmethylated CpG sequence in a test body with a sequence of bases in the specific DNA.

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BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows a gene maps of TLR9 knockout mice in the present invention and wild-type mice.

FIG. 2 shows the result of Southern blot analysis of TLR9 knockout mice in the present invention.

FIG. 3 shows the result of Northern blot analysis of spleen cells from TLR9 knockout mice in the present invention.

FIG. 4 shows the result of comparing the sequence of amino acids from TLR9 knockout mice in the present invention and the sequence of amino acids from wild-type mice.

FIG. 5 shows the result of measurement of TNF α , IL-6 or IL-12 production induced by CpG ODN, PGN or LPS in TLR9 knockout mice in the present invention and in wild-type mice.

FIG. 6 shows the result of cellular proliferation response induced by CpG ODN or LPS in TLR9 knockout mice in the present invention or in wild-type mice.

FIG. 7 shows the result of measurement of IL-12 production induced by CpG ODN or LPS in TLR9 knockout mice in the present invention or in wild-type mice.

FIG. 8 shows the result of expression of CD40, CD80, CD86, and MHC class II induced by CpG ODN or LPS in TLR9 knockout mice in the present invention and in wild-type mice.

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FIG. 9 shows the result of activation of NF- κ B induced by CpG ODN or LPS in TLR9 knockout mice in the present invention or in wild-type mice.

FIG. 10 shows the result of activation of JNK
5 induced by CpG ODN or LPS in TLR9 knockout mice in the present invention or in wild-type mice.

FIG. 11 shows the result of activation of IRAK induced by CpG ODN or LPS in TLR9 knockout mice in the present invention or in wild-type mice.

10 BEST MODE TO CARRY OUT THE PRESENT INVENTION

As bacterial DNA comprising an unmethylated CpG sequence in the present invention, any DNA derived from bacteria such as an oligodeoxynucleotide having an unmethylated CpG motif which activates immune cells such as T-cells, B-cells and antigen-presenting cells, and induces immune response can be used such as DNA derived from bacteria including *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Salmonella Typhimurium*, *Serratia marcescens*, *Shigella flexneri*, *Vibrio cholerae*, *Salmonella Minnesota*, *Porphyromonas gingivalis*, *Staphylococcus aureus*, *Corynebacterium diphtheriae*, *Nocardia coeliaca*, *Streptococcus pneumoniae*.

As a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG motif, there are no particular restrictions as long as the protein can specifically recognize bacterial DNA with an unmethylated CpG sequence, and can be exemplified by human-derived TLR9 shown in Seq. ID No. 2 in the list of sequence, a protein which comprises a sequence of amino acids wherein one or more of amino acids are deleted, substituted, or added in a sequence of amino acids shown in Seq. ID No: 2, and which specifically recognizes bacterial DNA having the unmethylated CpG sequence, or their recombinant proteins. The receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence can be prepared by well known methods based on the information of the DNA sequence and others.

DNA encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence of the present invention includes DNA encoding human-derived TLR9 shown in Seq. ID No: 2 in the list of sequence such as the one shown in Seq. ID No: 1, DNA comprising a sequence of amino acids wherein one or more of amino acids are deleted, substituted or added in a sequence of amino acids shown in Seq. ID No: 2, and which can specifically recognize bacterial DNA having the unmethylated CpG sequence mentioned above, or DNA hybridized with the DNA under stringent conditions and encoding a protein that can specifically recognize bacterial DNA having the unmethylated CpG sequence mentioned above. These can be prepared by well known methods based on the information of DNA sequence such as mouse RAW264.7 cDNA library or 129/SvJ mouse gene library for mouse-derived TLR9.

Further, it is possible to obtain DNA encoding a receptor protein specifically recognizing bacterial DNA having an immune-inducing unmethylated CpG sequence which has the same effect as TLR9, a receptor protein, by hybridizing mouse-derived DNA library

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with part or whole of a sequence of bases shown in SEQ ID No: 1 or its complementary sequence under stringent conditions to isolate the DNA hybridized with the probe. Conditions on hybridization to obtain the DNA can, for example, be hybridization at 42°C and wash treatment at 42°C with a buffer containing 1% x SSC and 0.1% of SDS, and more preferably be hybridization at 65°C and wash treatment at 65°C with a buffer containing 0.1 x SSC and 0.1% of SDS. Furthermore, beside the temperature conditions mentioned above, there are various factors effecting the stringency of hybridization, and it is possible for a person skilled in the art to realize the stringency equivalent to the stringency of hybridization illustrated above.

A fusion protein in the present invention can be the one obtained by combining a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence derived from mouse, human, and others with a marker protein and/or a peptide tag. A marker protein can be any marker protein previously well known, and can be exemplified by alkaline phosphatase, Fc region of an antibody, HRP, GFP and others. As a peptide tag in the present invention, it can be concretely exemplified by previously well-known peptide tags such as Myc tag, His tag, FLAG tag, GST tag. The fusion protein can be produced by a normal method, and is useful in purifying a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence by using affinity of Ni-NTA and His tag, detecting a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence, measuring of the amount of antibodies against a receptor protein specifically recognizing bacterial DNA

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having an unmethylated CpG sequence and as a research reagent in other relevant fields.

As an antibody which specifically binds to a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence in the present invention, it can be concretely exemplified by immune-specific antibodies such as a monoclonal antibody, a polyclonal antibody, a chimeric antibody, a single-chain antibody, a humanized antibody. These antibodies can be produced by a normal method by using a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence mentioned above as an antigen, and a monoclonal antibody is preferable in its specificity among them. The antibody which specifically binds to a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence such as a monoclonal antibody and others is useful, for example, in diagnosing diseases caused by the mutation or deletion of TLR9 or elucidating the molecular mechanism controlling TLR9.

An antibody against a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence can be produced by administering a fragment containing a receptor protein or an epitope specifically recognizing bacterial DNA having the unmethylated CpG sequence in animals (preferably, non-human), or a cell expressing the protein on the surface of its membrane by a conventional protocol, and any method can be used such as hybridoma method (Nature 256, 495-497, 1975), trioma method, human B cell hybridoma method (Immunology Today 4, 72, 1983), and EBV-hybridoma method (MONOCLONAL ANTIBODIES AND CANCER THERAPY, 77-96, Alan R. Liss, Inc. 1985), which are used for preparing monoclonal antibodies

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and bring an antibody produced by the cultured successive cell lines. The following explains a method of producing a monoclonal antibody which specifically binds to mouse-derived TLR9, that is, an mTLR9 monoclonal antibody, with
5 mouse-derived TLR9 as an example of a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence.

The mTLR9 monoclonal antibody can be produced by a normal method of culturing hybridoma producing mTLR9
10 monoclonal antibody in vivo or in vitro. For example, in an in vivo systems, the monoclonal antibody can be obtained by culturing in the visceral cavity of rodents, preferably of mice or rats, and in an in vitro system, the monoclonal antibody can be obtained by culturing in a medium for
15 culturing animal cells. A medium used for culturing hybridoma in an in vitro system can be exemplified by cell culture media such as RPMI1640 or MEN and others comprising antibiotics such as streptomycin or penicillin.

The hybridoma producing mTLR9 monoclonal antibody
20 can be produced by immunizing BALB/c mouse with TLR9, a receptor protein obtained from mouse and others, fusing a spleen cell from an immunized mouse and a mouse NS-1 cell (ATCC TIB-18) by a normal method, and screening them by immunofluorescence staining patterns. A method of
25 separating/isolating the monoclonal antibody can be any one as long as it is a method usually used for purifying proteins, and liquid chromatography such as affinity chromatography and others can be a concrete example.

It is also possible to apply the method of a
30 single-chain antibody (US Patent No. 4946778) to produce single-chain antibodies against receptor proteins

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specifically recognizing bacterial DNA having the above-mentioned unmethylated CpG sequence of the present invention. Further, it is possible to use transgenic mice or other mammals and the like to

express humanized antibodies, isolate/identify the clones expressing a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence by using the antibodies, and purify the polypeptides by affinity chromatography. The antibodies against receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence are useful in elucidating the molecular mechanism of a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence.

It is also possible to carry out a functional analysis of a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence by using fusion proteins obtained by fusing proteins labeled with fluorescent substances such as FITC (fluorescein isothiocyanate) or tetramethylrhodamine isocyanate, fusion proteins labeled with radio isotopes such as ^{125}I , ^{32}P , ^{35}S or ^3H , enzymes such as Alkaline phosphatase, peroxidase, β -Galactosidase or Phycoerythrin, or fluorescent proteins such as Green Fluorescent Protein (GFP). A method of immunoassay can be exemplified by RIA, ELISA, fluorescence antibody method, plaque forming cell assay, spot method, hemagglutination reaction method, Ouchterlony Method, and others.

The present invention relates to a host cell comprising an expressing system that can express a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence. Introduction of a gene encoding a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence into a host cell can be carried out by the methods described in a number of standard laboratory manuals such as in Davis et al. (BASIC METHODS IN MOLECULAR BIOLOGY, 1986) and Sambrook et al. (MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N. Y., 1989), such as calcium phosphate transfection, DEAE-dextran-mediated transfection, transvection, microinjection, cationic lipid-mediated transfection, electroporation, transduction, scrape loading, ballistic introduction, infection and others. A host cell can be exemplified by bacterial prokaryotes such as *Escherichia coli*, *Streptomyces*, *Bacillus subtilis*, *Streptococcus*, *Staphylococcus* and others, fungal cells such as yeast and *Aspergillus*, insect cells such as *Drosophila* S2 or *Spodoptera Sf9* and others, and animal and plant cells such as L cell, CHO cell, COS cell, Hela cell, C127 cell, BALB/c3T3 cell (including mutant strains lacking dihydrofolate reductase, thymidine kinase or others), BHK 21 cell, HEK293 cell, Bowes Melanoma cell, oocytes, and others.

Further, the expression system can be any one as long as it is a system that can express

a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence in a host cell, and can be exemplified by expression systems derived from chromosome, episome and virus, such as vectors derived from bacterial plasmid, yeast plasmid, papovavirus such as SV40, vaccinia virus, adeno virus, fowl poxvirus, pseudorabies virus, or vectors derived from retrovirus, vectors derived from bacteriophage or transposon or their combinations, which can be exemplified by plasmids such as cosmid and phagemid, which are derived from genetic factors of plasmids and bacteriophage. These expressing systems may comprise a control sequence that not only causes expression but also regulates expression.

A receptor protein specifically recognizing a host cell comprising the expressing system or a cell membrane of the cell, bacterial DNA comprising an unmethylated CpG sequence obtained by culturing, and the cell can be used for the screening methods of the present invention as mentioned below. For example, a method described in F. Pietri-Rouxel et al. (Eur. J. Biochem., 247, 1174-1179, 1997) can be used as a method for obtaining cell membrane, and well known methods including ammonium sulfate or ethanol precipitation, acid extraction, anion- or cation-exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxyapatite chromatography and lectin chromatography, preferably high-performance liquid chromatography can be used to collect a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence from the cell culture and to purify it. Specifically, it is possible to obtain a receptor protein specifically recognizing the bacterial DNA having an unmethylated CpG sequence by using a column to which a receptor protein antibody specifically recognizing bacterial DNA having the anti-unmethylated CpG sequence of anti-TLR9 monoclonal antibodies and others is bound, or in case an ordinary peptide tag is bound to a receptor protein such as TLR9 etc. specifically recognizing a column to which a substance having an affinity with a peptide tag is bound for affinity chromatography.

A non-human animal excessively expressing a gene encoding a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence mentioned above in the present invention can be a non-human animal producing a large amount of receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence compared with wild-type non-human animals. Further, a non-human animal whose gene function encoding a receptor protein specifically recognizing bacterial DNA

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having an unmethylated CpG sequence is deleted on the chromosome is a non-human animal wherein part or whole of genes encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence on the a chromosome are inactivated by genetic mutations such as damaged, deleted, substituted, and others, and which lost a function of expressing a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence. Although the non-human animal used in the present invention can be exemplified by a non-human animal including rodents such as rabbits, mice, rats and others, it is not restricted to the animals.

Further, refractory against bacterial DNA having an unmethylated CpG sequence in the present invention means that the reactivity against stimuli by bacterial DNA shown by an organism, or a cell, a tissue or an organ constituting the organism is declined or almost totally lost. Therefore, a non-human animal with refractory against bacterial DNA having an unmethylated CpG sequence in the present invention is a non-human animal such as mice, rats, or rabbits, wherein the an organism's reactivity against bacterial DNA, or a cell, a tissue or an organ constituting the organism is declined or almost totally lost. Further, stimuli by bacterial DNA can be exemplified by an in vivo stimulus caused by administrating bacterial DNA to an organism, or an in vitro stimulus caused by contacting cells separated from an organism with bacterial DNA. Concretely, a non-human animal such as TLR9 knockout mice wherein TLR9 gene functions are destroyed on the chromosome can be an example.

Homozygote non-human animals born following Mendel's Law include mice deficient of or excessively expressing receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence and their wild-type littermates, and it is preferable to use wild-type non-human animals, that is, the same kind of animal as a non-human animal wherein gene functions encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence are destroyed or are excessive, more preferably their littermate animals, for example, during the screening of the present invention described below because accurate comparative experiments can be carried out at the level of individuals by using the homozygote non-human animals with its receptor proteins destroyed or the one with receptor proteins expressing excessively or the wild-type non-human animals born from the same mother at the same time. In the following, a method of producing non-human animals wherein gene functions encoding a receptor protein specifically recognizing bacterial DNA having the unmethylated CpG sequence are destroyed or excessively

expressed on the chromosome is explained using knockout mice or transgenic mice whose receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence as an example.

For example, as for a mouse wherein gene functions encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence are destroyed on a chromosome such as TLR9, that is, a knockout mouse lacking receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence, gene fragments obtained from mouse gene library by a method of PCR or the like are used to screen genes encoding receptor proteins specifically recognizing bacterial DNA having the unmethylated CpG sequence, subclone a gene encoding a receptor protein specifically recognizing bacterial DNA having the screened unmethylated CpG sequence with viral vectors and others, and specified by DNA sequencing. Whole or part of the gene in the clone encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is substituted with pMC1 neo gene cassette and others, and a targeting vector is produced by introducing diphtheria toxin A fragments (DT-A) genes or herpes simplex virus thymidine kinase (HSV-tk) genes and others on 3'-end side.

The produced targeting vector is linearized, introduced into ES cells by electroporation method and others, homologous recombination is performed, and ES cells which have caused homologous recombination by antibiotics such as G418 or gancyclovir (GANC) and others are selected from the homologous recombinants. It is preferable to confirm by Southern blot technique that the selected ES cells are targeted recombinants. The clones of the confirmed ES cells are introduced to mouse blastocysts by microinjection, and the blastocysts are returned to recipient mice, and chimera mice were produced. The chimera mouse was intercrossed with a wild-type mouse to produce a heterozygote mouse, and the heterozygote mice are intercrossed to produce a knockout mouse lacking a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence in the present invention. Further, a method of confirming whether knockout mice lacking a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence is obtained, for example, may be examined by Northern blot technique, which isolates RNA from the mouse obtained by the method mentioned above, or the expression in the mice may be examined by Western blot technique.

The fact that the produced TLR9 knockout mouse is refractory against bacterial DNA

having an unmethylated CpG sequence can be confirmed by measuring the levels of the production of TNF- α , IL-6, IL-12, IFN- γ and others in the cells whose CpG ODN was contacted in vivo or in vitro with immune cells such as macrophages, mononuclear cells, dendritic cells from TLR9 knockout mice, the proliferation of response of spleen B cells, the expression of antibodies such as CD40, CD80, CD86, MHC class II on the surface of spleen B cells, and the activation of molecules on the signal transduction pathway of NF- κ B, JNK, IRAK and others. The knockout mice lacking TLR9 in the present invention can be used to elucidate functional mechanisms of bacterial DNA and others having an unmethylated CpG sequence and to developing vaccine against bacterial infections.

Transgenic mice lacking receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence can be generated by constructing introduced genes by fusing chicken β actin, mouse neurofilament, promoters such as SV40, and rabbit β -globin, polyA such as SV40 or intron with cDNA encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence such as TLR9, microinjecting the introduced genes to pronucleus of mouse fertilized eggs, transplanting the obtained cells to an oviduct of recipient mice after culturing them, then breeding the transplanted animals, and selecting child mice having the cDNA from born child mice. Further, selection of the child mice having cDNA can be performed by dot hybridization wherein crude cDNA was extracted from mouse tails and others, and genes encoding receptor proteins specifically recognizing bacterial DNA having an introduced unmethylated CpG sequence is used as a probe, or PCR method using specific primers and others.

Further, the use of whole or part of DNA encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence in the present invention enables us to prepare cells effective for genetic treatments for diseases caused by the deletion or abnormality of receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence. Methods of preparing the cells in the present invention can be exemplified by a method wherein part or whole of the DNA in the present invention is introduced into cells lacking gene functions encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence on the chromosome by transfection and others, and thus obtaining a cell expressing receptor proteins specifically recognizing bacterial DNA having the unmethylated CpG sequence. It is preferable to use a cell in which the DNA and others is integrated onto the chromosome and shows TLR9

activity in a stable manner, particularly as a cell expressing receptor proteins specifically recognizing bacterial DNA having the unmethylated CpG sequence.

Furthermore, the use of DNA encoding receptor proteins specifically recognizing bacterial DNA having the unmethylated CpG sequence, antibodies against receptor proteins specifically recognizing bacterial DNA having a fused unmethylated CpG sequence comprising a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence bound to a marker protein and/or a peptide tag, a host cell comprising an expression system which can express a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence, non-human animals excessively expressing genes encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence, non-human animals lacking gene functions encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence on a chromosome, cells expressing receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence enables us to screen agonists or antagonists of the receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence in the present invention, or suppressing or promoting substances reactive to bacterial DNA having an unmethylated CpG sequence. What is obtained by the screening may be suppressing or promoting substances against bacterial infected diseases, suppressing agents, preventing agents or remedies against allergic diseases or cancers, agents suppressing or promoting side effects in genetic therapy or the like, or substances useful for diagnosing/treating diseases or the like caused by the deletion or abnormality of TLR9 activity.

Although the TLR activities can concretely be exemplified by a function of reacting specifically to bacterial DNA having an unmethylated CpG sequence and transmitting signals into cells, and a signal transduction function is a function of producing cytokines such as $\text{TNF-}\alpha$, IL-6, IL-12, IFN- γ or the like, a function of producing nitrous acid ion, a function of proliferating cells, a function of expressing antibodies such as CD40, CD80, CD86, MHC class II and others on the surface of cells, and a function of activating molecules in signal transduction pathway of TLR9 such as NF- κ B, JNK, IRAK and others, it is not limited to these functions.

A screening method of agonists or antagonists of receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence in the present invention

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can concretely be exemplified by a method of performing
in vitro culture of immune cells such as macrophages, spleen
cells or dendritic cells, cells expressing a receptor
protein specifically recognizing bacterial DNA having an
5 unmethylated CpG sequence, cells expressing a protein having
reactivity against bacterial DNA having an unmethylated CpG
sequence in a cell expressing a receptor protein
specifically recognizing bacterial DNA having an
unmethylated CpG sequence under the presence of target
10 substance, and measuring/evaluating TLR9 activities, or a
method of administrating target substance to wild-type non-
human animals, non-human animals lacking a gene function of
encoding receptor proteins specifically recognizing
bacterial DNA having an unmethylated CpG sequence, or non-
15 human animals excessively expressing genes encoding receptor
proteins specifically recognizing bacterial DNA having an
unmethylated CpG sequence, and measuring/evaluating TLR9
activities of immune cells such as macrophages, spleen cells
or dendritic cells derived from these non-human animals.
20 The TLR9 activities are compared with corresponding TLR9
activities of the wild-type non-human animals of the same
species as the transgenic non-human animals, and when a
difference of the TLR9 activities between the wild-type non-
human animals and the non-human animals lacking the gene
25 function of encoding receptor proteins specifically
recognizing bacterial DNA having the unmethylated CpG
sequence is larger than that of a case in which no substance
is administered, then that is an indication that the target
substance is the agonist, and when the difference is smaller,
30 then that is an indication that the target substance is the
antagonist.

Further, in evaluating and measuring the levels of
macrophage activities or spleen cell activities, it is

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preferable to evaluate and compare them with the measurement values obtained from wild-type non-human animals, especially wild-type non-human animals born from the same parent to remove variances arising from individual differences. The same also applies to screening of suppressing or promoting substances reactive to bacterial DNA having an unmethylated CpG sequence shown below.

Screening methods for suppressing or promoting substances reactive to bacterial DNA having an unmethylated CpG sequence can concretely be exemplified by a method comprising the steps of carrying out in vitro incubation of proteins or cell membranes expressing the proteins having a reactivity against bacterial DNA having an unmethylated CpG sequence in the presence of target substances and bacterial DNA having an unmethylated CpG sequence, measuring/evaluating the reactivity of the protein, or a method comprising the steps of first making macrophages or spleen cells obtained from non-human animals whose gene functions encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence are destroyed on a chromosome contact in vitro with target substances, then culturing the macrophages or spleen cells in the presence of bacterial DNA having an unmethylated CpG sequence, and measuring/evaluating the levels of macrophage activities shown by the macrophages or the levels of spleen cell activities shown by the spleen cells, a method comprising the steps of making macrophages or spleen cells

obtained from non-human animals whose gene functions encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence contact in vitro with bacterial DNA having an unmethylated CpG sequence, then culturing the macrophages or spleen cells in the presence of target substances, and measuring/evaluating the levels of macrophage activities shown by the macrophages or the levels of spleen cell activities shown by the spleen cells, and a method of comprising the steps of first administering target substances to non-human animals whose gene functions encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence on a chromosome first, then culturing the macrophages or spleen cells obtained from the non-human animals in the presence of bacterial DNA having an unmethylated CpG sequence, and measuring/evaluating the levels of macrophage activities shown by the macrophages or the levels of spleen cell activities shown by the spleen cells, a method comprising the steps of first administering target substances to non-human animals whose gene functions encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence is destroyed on a chromosome, then infecting the non-human animals by bacteria, and measuring/evaluating the levels of macrophage activities shown by macrophages or the levels of spleen cell activities shown by the spleen cells obtained from non-human animals, a method of the steps of first administering target substance to non-human animals whose gene functions encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence on a chromosome, and measuring/evaluating the levels of macrophage activities shown by macrophages or the levels of spleen cell activities shown by spleen cells obtained from the non-human animals, a method comprising the steps of first infecting with bacteria non-human animals whose gene functions encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence are destroyed on a chromosome, then culturing macrophages or spleen cells obtained from the non-human animals in the presence of target substances, and measuring/evaluating the levels of macrophage activities shown by macrophages or the levels of spleen cell activities shown by spleen cells obtained from the non-human animals, a method comprising the steps of administering target substances to non-human animals whose gene functions are encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence are destroyed, infecting the non-human animals by bacteria, and measuring/evaluating the levels of macrophage activities or spleen cell activities in the non-human animals, and a method comprising the steps of infecting non-

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human animals whose gene functions encoding proteins having reactivity against bacterial DNA having an unmethylated CpG sequence are destroyed on a chromosome first, then administering the target substances to the non-human animals, and measuring/evaluating the levels of macrophage activities or spleen cell activities in the non-human animals. Although as bacterial DNA having an unmethylated CpG sequence used in the screening methods, it is preferable to use CpG ODN (TCC-ATG-ACG-TTC-CTG-ATG-CT: SEQ ID No: 5), it is not limited to this.

The present invention also relates to a kit used to diagnose diseases relating to the activity or expression of receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence by comparing a sequence of DNA encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence in a test body with a sequence of DNA encoding receptor proteins specifically recognizing bacterial DNA having a unmethylated CpG sequence in the present invention. The detection of mutated DNA encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence can be carried out by detecting genetically mutated individuals at the level of DNA, and is effective for diagnosing diseases caused by hypotypic expression, hypertypic expression or mutated expression of receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence. Although a test body used in the detection can concretely be exemplified by genomic DNA of cells from subjects obtainable by biopsy from blood, urine, saliva, tissue and others, RNA, or cDNA, it is not limited to these. In using the test body, it is possible to use the ones amplified by PCR and others. The deficiency or insertional mutation in sequences of bases can be detected

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by the changes of amplified products in size compared with normal genes, and point mutation can be identified by hybridizing the amplified DNA with the gene encoding receptor proteins specifically recognizing bacterial DNA having labelled unmethylated CpG sequence. It is possible to diagnose or conclude diseases relevant to activity or expression of receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence by detecting mutation of a gene encoding receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence. The kit may include reagents useful for performing PCR of the genomic DNA, RNA or cDNA of the cells from the test subjects.

The present invention also relates to a probe diagnosing a disease related to activities or expressions of a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence comprising whole or part of antisense chain of DNA or RNA encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence, and a kit used to diagnose diseases relating to activities or expressions of a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence comprising an antibody specifically bound to a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence of the probe and/or in the present invention. A probe used for the diagnosis is whole or part of an antisense chain of DNA (cDNA) or RNA (cRNA) encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence, and there is no limitations on the probe as long as it is long enough (at least 20 bases or more) to establish as a probe. In order to make an antibody specifically bound to a receptor

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protein specifically recognizing bacterial DNA having an unmethylated CpG sequence of the probe and/or in the present invention an active component of a medicine diagnosing diseases such as bacterial infection and others, it is
5 preferable to dissolve it into appropriate buffers or sterilized water in which a probe is not decomposed. Further, it is possible to use the clinical test pharmaceuticals to diagnose a patient's symptoms such as bacterial infection diseases and others in the ways such as
10 immunofluorescence (Dev. Biol. 170, 207-222, 1995, J. Neurobiol. 29, 1-17, 1996), In situ hybridization (J. Neurobiol. 29, 1-17, 1996), or in situ PCR or others.

A pharmaceutical composition of the present invention can be any one as long as it comprises whole or
15 part of the receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence of TLR9 or others, or comprises an agonist or an antagonist of the receptor protein. The pharmaceutical composition also contains a pharmaceutically acceptable diluent and
20 optionally other additives. Concretely, vaccines against bacterial infectious diseases, vaccines against cancers, treating medicine for patients having allergies such as bronchial asthma, reversal agents, suppressing agents, inhibiting agents and others for side effects by the
25 existence of a CpG motif inhibiting genetic treatments or treatments using antisenseoligonucleotides can be exemplified.

As mentioned above, a kit testing diagnoses relevant to the deletion, substitution and/or addition of
30 DNA sequence encoding a receptor proteins specifically recognizing bacterial DNA having an unmethylated CpG sequence of the present invention can be any one as long as

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it comprises DNA encoding TLR9, and comparing a sequence of
bases of DNA encoding the TLR9 with a sequence of bases of
DNA encoding a receptor protein specifically recognizing
bacterial DNA having an unmethylated CpG sequence in a
5 tested body enables us to diagnose diseases related to
deletion, substitution and/or addition of DNA sequence

encoding a receptor protein specifically recognizing bacterial DNA having an unmethylated CpG sequence such as cancer, allergy, infectious diseases and others.

In the following, the present invention will further be explained with concrete examples. However, the technical scope of the present invention is not limited in any way by the following examples.

Example 1: Cloning of TLR9

As a result of a GenBank search using the information of DNA sequence of human TLR4, a mouse EST having a significant homology (Registration No. AA273731; mouse) was found. Using PCR amplified mouse EST as a probe, mouse RAW 264.7 cDNA library was screened and a full length cDNA clone shown in Seq. ID No. 3 comprising the complete TLR9 open reading frame was isolated. Performing a GenBank search based on the information of DNA sequence of the mouse TLR9, a human genomic sequence having a high level of homology was found. Based on the human genomic sequence, cDNA ends were amplified to isolate cDNA of the full length human TLR9 having a sequence of bases in Seq. ID No. 1 from U937 cells (J. Immunol. 163, 5039-5048, 1999).

Example 2: Production of TLR knockout mice

The TLR9 genomic DNA was isolated from 129/SvJ mouse genomic library (Stratagene), subcloned in pBluescript II SK(+) vectors (Stratagene), and characterized by restriction enzyme mapping and DNA sequencing analysis. The targeting vector was constructed by replacing a 1.0 kb fragment encoding part of LRR (leucine-rich repeat) region with a neomycin-resistance gene cassette (pMC1-neo; Stratagene), and a herpes simplex virus thymidine kinase (HSV-TK) was inserted for negative selection (Fig. 1). The targeting vector was linearized, and was electroporated into embryonic stem cells (ES cells) of E14-1, then 292 pieces of clones showing G418 and gancyclovir resistance were selected, and 14 pieces of clones were screened by PCR and Southern blotting.

Chimeric mice were produced by microinjecting 3 pieces of targeted ES clones comprising mutated TLR9 allele into C57BL/6 mouse blastocysts. The male chimeric mice were intercrossed with C57BL/6 female mice to produce a heterozygote F1 mouse, and a homozygote mice (TLR9 knockout mouse: TLR9^{-/-}) was obtained by intercrossing heterozygote F1 mouse (Fig. 2). To confirm that the obtained mouse was homozygote,

various genomic DNA extracted from a mouse tail was digested by ScaI to perform Southern blotting using the probe shown in Fig. 1. The TLR9 knockout mice (TLR9^{-/-}) of the present invention were produced following Mendel's law, and had not shown remarkable abnormality for 12 weeks.

To confirm that the inactivation of TLR9 arises by mutation, total RNA (10 μ g) extracted from spleen cells from wild-type mice (+/+) and TLR9 knockout mice (-/-) was electrophoresed, and transferred to nylon membranes, Northern blotting was performed with the use of cDNA specific to TLR9 c-terminal fragments or N-terminal fragments labeled with [³²P], or β -actin (Fig. 3). The result shows that N-terminal fragments of TLR9 mRNA were not detected from the spleen cells of TLR9 knockout mice. Further, with a C-terminal fragment as a probe, almost the same size of Tlr9 transcripts derived from mutated mice as the ones from wild-type mice were detected. However, the amount of the production was small. Then, RT-PCR was performed using mRNA of spleen cells obtained from mutated mice to sequence the obtained products. The result shows that the Tlr9 gene transcript comprises neo gene, and stop codons appear in a N-terminal domain of TLR9 by inserting the neo, and functional TLR9 proteins does not appear in mutated mice (Fig. 4). Further, as a result of examining lymph cells from TLR9 by flowcytometry knockout mice, no abnormal compositions were found.

Example 3: Preparation of peritoneal macrophages

2ml of 4% thioglycolic acid medium (DIFCO) was injected to each peritoneum of wild-type mice and TLR9 knockout mice (TLR9^{-/-}), peritoneal exudation cells were isolated from peritonea from each mouse after 3 days, the cells were cultured in RPMI1640 medium to which 10% of fetal bovine serum (GIBCO) was added at 37°C for 2 hours, and remove the unattached cells by washing with ice-chilled Hank's buffered salt solution (HBSS; GIBCO), and the attached cells were used as peritoneal macrophages in the following experiments.

Experiment 4: Response to bacterial DNA having an unmethylated CpG sequence in TLR9 knockout mice

It has recently been shown that the response of CpG ODN (oligodeoxynucleotide) is dependent on MyD88, an adapter protein in a signaling transduction pathway mediating TLR. Although the MyD88 knockout mice do not show response to CpG ODN, TLR2 knockout

mice or TLR4 knockout mice show normal response to it. This shows that CpG ODN recognizes TLRs other than TLR2 and TLR4, and then the response of a TLR9 knockout mouse against CpG ODN was examined. First, the amount of producing inflammatory cytokines in peritoneal macrophages were measured in the following way.

The macrophages prepared in Example 3 are co-cultured with various concentrations of CpG ODN shown in Fig. 5 (0.1 or 1.0 μ M; TIB MOLBIOL; TCC-ATG-ACG-TTC-CTG-ATG-CT), PGN (10 μ g/ml; Sigma and Fluka; derived from *Staphylococcus aureus*), LPS (1.0 μ g/ml; Sigma; derived from *Salmonella minnesota* Re-595) in the presence or absence of $\text{INF } \gamma$ (30 unit/ml). The concentrations of $\text{TNF } \alpha$, IL-6 and IL-12 p40 in the supernatants after culturing were measured by ELISA, and the results are shown in Fig. 5. The results show that the macrophages from wild-type mice (Wild-type) produce $\text{TNF } \alpha$, IL-6 and IL-12 in response to CpG ODN, and further stimulation by $\text{INF } \gamma$ and CpG ODN increases the amount of producing $\text{TNF } \alpha$, IL-6 and IL-12. However, the macrophages derived from TLR9 knockout mice ($\text{TLR9}^{-/-}$) did not produce a detectable level of inflammatory cytokines in response to CpG ODN even in the presence of $\text{INF } \gamma$. Further, it was found that the macrophages derived from wild-type mice and TLR9 knockout mice produce almost the same level of $\text{TNF } \alpha$, IL-6 and IL-12 in response to LPS or PGN (Fig. 5). Each experimental result shows the average level of $n=3$. N.D. in the figures means not detected.

Response of spleen cells from wild-type mice (Wild-type) and TLR9 knockout mice ($\text{TLR9}^{-/-}$) against CpG ODN or LPS was also examined. The spleen cells from each mouse (1×10^5) were isolated to culture in 96 well plates by CpG DNA or LPS of various concentrations shown in Fig. 6, and the spleen cells were stimulated. 40 hours later from culturing, 1 μ Ci of [^3H]-thymidine (Dupont) was added, and then further cultured for 8 hours. The amount of uptaking [^3H]-thymidine was measured by β scintillation counter (Packard) (Fig. 6). The results that although the spleen cells from wild-type mice promote cell proliferating reactions depending on the amount of administering CpG ODN or LPS, the spleen cells from TLR9 knockout mice did not show any cell proliferating reaction by CpG ODN even with the stimulus of any concentration of CpG ODN. Further, the amount of expressing Major Histocompatibility Complex (MHC) class II on the surface of B cells derived from wild-type mice in response to CpG ODN was increased. However, such increase of the amount of expressing MHC class II induced by CpG ODN in B cells derived

from TLR9 knockout mice was not observed. These facts show that the macrophages or B cells from TLR9 knockout mice specifically lack the response against CpG ODN.

Next, it is well known that DNA derived from bacteria comprising CpG ODN potentially stimulates dendritic cells, and supports the development of Th1 cell (EMBO J. 18, 6973-6982, 1999, J. Immunol. 161, 3042-3049, 1998, Proc. Natl. Acad. Sci. USA 96, 9305-9310, 1999). Then, the production of CpG ODN-inducing cytokines and the upregulation of the surface molecule of dendritic cells derived from bone marrow were examined. The bone marrow cells from wild-type mice (Wild-type) or TLR9 knockout mice (TLR9^{-/-}) were cultured with 10ng/ml mouse granulocyte macrophage-colony stimulating factor (Peprotech) in RPMI1640 medium supplemented with 10% fetal bovine serum (J. Exp. Med. 176, 1693-1702, 1992), at day 6 of the culture, immature dendritic cells were harvested and cultured in the presence or absence of 0.1 μ M CpG ODN or 0.1 μ g/ml LPS in RPMI1640 medium supplemented with 10% fetal bovine serum for 2 days. After the culture, the concentration of IL-12 p40 in the supernatants was measured by ELISA (Fig. 7). The result shows that the dendritic cells derived from wild-type mice produced IL-12 in response to CpG ODN while the dendritic cells derived from TLR9 knockout mice did not induce the production of IL-12 in response to CpG ODN.

After culturing in RPMI supplemented with 10% fetal bovine serum was cultured which contains 10ng/ml mouse granulocyte macrophage-colony stimulating factor (Peprotech), the dendritic cells harvested at day 6 were stained with biotinylated antibodies against CD40, CD80, CD86 or MHC class II, developed with streptavidine labeled with phycoerythrin (PE; PharMingen). The cells were examined by using a FACSCalibur with CELLQuest software (Becton Dickinson) (Fig. 8). The result shows that stimulation by CpG ODN promotes the expression of CD40, CD80, CD86 and MHC class II on the surface of dendritic cells derived from wild-type mouse while it does not promote the expression of these molecules on the surface of dendritic cells derived from TLR9 knockout mouse by the stimulation of CpG ODN (Fig. 8). The dendritic cells from wild-type mice and from TLR9 knockout mouse show similar responses in response to LPS. This result shows that TLR9 is a receptor essential for cell response to CpG ODN.

Example 5: activation of NF- κ B, JNK and IRAK in response to
CpG ODN of macrophages derived from TLR9 knockout mice

It is known that signaling via TLRs activates IRAK, a serine-threonine kinase mediated by MyD88, an adaptor molecule, and subsequently activates MAP kinase and NF- κ B (Immunity 11, 115-122, 1999). Whether CpG ODN activates the intracellular signaling or not was examined. The peritoneal macrophages (1×10^6 cells) from wild-type and TLR9^{-/-} mice in Example 3 were stimulated by $1.0 \mu\text{M}$ of CpG ODN or $1.0 \mu\text{g/ml}$ of LPS from *Salmonella minnesota* Re-595 for the periods indicated in Fig. 9, nucleoproteins were extracted from the macrophages obtained from each mouse to be incubated together with a specific probe comprising NF- κ B DNA-binding sites, electrophoresed, and then visualized by autoradiography (Fig. 9).

The result shows that when stimulated by CpG ODN, the macrophages derived from wild-type mice increased NF- κ B DNA-binding activity while the macrophages derived from TLR9 knockout mice did not increase NF- κ B DNA-binding activity. When stimulated by LPS, the macrophages derived from TLR9 knockout mice and the macrophages derived from the wild-type mice show similar NF- κ B activities.

The result shows that the macrophages derived from a TLR9 knockout mouse specifically lack NF- κ B activity by the induction of CpG ODN. The arrows in the figures indicate the sites of the compounds of NF- κ B and specific probes, and the arrowheads indicate the sites of specific probes only.

As shown above, the macrophages from wild-type mice and TLR9 knockout mice stimulated by CpG ODN or LPS for the periods indicated in Fig. 10 and Fig. 11 were dissolved into a solvent buffer (a buffer comprising 1.0% Triton X-100, 137mM of NaCl, 20mM of Tris-HCl, 5mM of EDTA, 10% glycerol, 1mM of PMSF, $20 \mu\text{g/ml}$ of aprotinin, $20 \mu\text{g/ml}$ of leupeptin, 1mM of Na_3VO_4 and 10mM of β -glycerophosphate at the final concentrations; pH8.0), the cell lysates were immunoprecipitated with anti-JNK antibody (Santa Cruz) or anti-IRAK antibody (Hayashibara Seikagaku Kenkyujo Kabushiki Kaisha). As described in a reference (Immunity 11, 115-122, 1999), the JNK activity and IRAK activity were measured by in vitro kinase assay using GST-c-Jun fusion protein (GST-c-Jun) as a substrate (top figures of Fig 10 and Fig.11; GST-c-Jun, Auto).

The cell lysates were separated by SDS-polyacrylamide gel electrophoresis to transfer them onto a nitrocellulose membrane and blotted the membrane with anti-JNK antibody (Santa Cruz) or anti-IRAK antibody (Transduction Laboratories) to visualize using an

enhanced chemiluminescent system (Dupont) (bottom figures of Fig. 10 and Fig. 11; WB). The result shows that CpG ODN activates JUN and IRAK of the macrophages derived from wild-type mice while it does not activate JUN and IRAK of the macrophages derived from TLR9 knockout mice (Fig. 10 and Fig.11). It is therefore found that the signaling transduction mediated by CpG ODN depends on TLR9.

INDUSTRIAL APPLICABILITY

Bacteria-derived DNA comprising an unmethylated CpG motif significantly activates immune cells and induce Th1 response, while a receptor recognizing such bacterial DNA remained unknown. The present invention has revealed a receptor of oligonucleotides comprising an unmethylated CpG sequence of bacterial DNA and will enable us to elucidate a receptor protein TLR9, a member of TLR family, specifically recognizing bacterial DNA having an unmethylated CpG sequence, the genetic DNA encoding it or others, which will be useful to diagnose and treat bacterial diseases and others. The use of the TLR9 knockout animals will also enable us to elucidate functional mechanisms of DNA derived from bacteria at the molecular level.

SEQUENCE LISTING

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					Met Gly Phe	
					1	

tgc cgc	agc gcc	ctg cac	ccg ctg	tct ctc	ctg gtg	cag gcc	atc atg	163
Cys Arg	Ser Ala	Leu His	Pro Leu	Ser Leu	Leu Val	Gln Ala	Ile Met	
5		10		15				

ctg gcc	atg acc	ctg gcc	ctg ggt	acc ttg	ccf gcc	ttc cta	ccc tgt	211
Leu Ala	Met Thr	Leu Ala	Leu Gly	Thr Leu	Pro Ala	Phe Leu	Pro Cys	
20		25		30		35		

gag ctc	cag ccc	cac ggc	ctg gtg	aac tgc	aac tgg	ctg ttc	ctg aag	259
Glu Leu	Gln Pro	His Gly	Leu Val	Asn Cys	Asn Trp	Leu Phe	Leu Lys	
	40		45		50			

tct gtg ccc cac ttc tcc atg gca gca ccc cgt ggc aat gtc acc agc	307
Ser Val Pro His Phe Ser Met Ala Ala Pro Arg Gly Asn Val Thr Ser	
55 60 65	
ctt tcc ttg tcc tcc aac cgc atc cac cac ctc cat gat tct gac ttt	355
Leu Ser Leu Ser Ser Asn Arg Ile His His Leu His Asp Ser Asp Phe	
70 75 80	
gcc cac ctg ccc agc ctg cgg cat ctc aac ctc aag tgg aac tgc ccg	403
Ala His Leu Pro Ser Leu Arg His Leu Asn Leu Lys Trp Asn Cys Pro	
85 90 95	
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Pro Val Gly Leu Ser Pro Met His Phe Pro Cys His Met Thr Ile Glu	
100 105 110 115	
ccc agc acc ttc ttg gct gtg ccc acc ctg gaa gag cta aac ctg agc	499
Pro Ser Thr Phe Leu Ala Val Pro Thr Leu Glu Glu Leu Asn Leu Ser	
120 125 130	
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Tyr Asn Asn Ile Met Thr Val Pro Ala Leu Pro Lys Ser Leu Ile Ser	
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Leu Ser Leu Ser His Thr Asn Ile Leu Met Leu Asp Ser Ala Ser Leu	
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Thr Ala Leu Arg Val Leu Asp Val Gly Gly Asn Cys Arg Arg Cys Asp			
245	250	255	
cac gct ccc aac ccc tgc atg gag tgc cct cgt cac ttc ccc cag cta			931
His Ala Pro Asn Pro Cys Met Glu Cys Pro Arg His Phe Pro Gln Leu			
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cat ccc gat acc ttc agc cac ctg agc cgt ctt gaa ggc ctg gtc ttg			979
His Pro Asp Thr Phe Ser His Leu Ser Arg Leu Glu Gly Leu Val Leu			
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aag gac agt tct ctc tcc tgg ctg aat gcc agt tgg ttc cgt ggg ctg			1027
Lys Asp Ser Ser Leu Ser Trp Leu Asn Ala Ser Trp Phe Arg Gly Leu			
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gga aac ctc cga gtg ctg gac ctg agt gag aac ttc ctc tac aaa tgc			1075
Gly Asn Leu Arg Val Leu Asp Leu Ser Glu Asn Phe Leu Tyr Lys Cys			
310	315	320	
atc act aaa acc aag gcc ttc cag ggc cta aca cag ctg cgc aag ctt			1123
Ile Thr Lys Thr Lys Ala Phe Gln Gly Leu Thr Gln Leu Arg Lys Leu			
325	330	335	
aac ctg tcc ttc aat tac caa aag agg gtg tcc ttt gcc cac ctg tct			1171
Asn Leu Ser Phe Asn Tyr Gln Lys Arg Val Ser Phe Ala His Leu Ser			
340	345	350	355
ctg gcc cct tcc ttc ggg agc ctg gtc gcc ctg aag gag ctg gac atg			1219
Leu Ala Pro Ser Phe Gly Ser Leu Val Ala Leu Lys Glu Leu Asp Met			
360	365	370	
cac ggc atc ttc ttc cgc tca ctc gat gag acc acg ctc cgg cca ctg			1267
His Gly Ile Phe Phe Arg Ser Leu Asp Glu Thr Thr Leu Arg Pro Leu			
375	380	385	
gcc cgc ctg ccc atg ctc cag act ctg cgt ctg cag atg aac ttc atc			1315
Ala Arg Leu Pro Met Leu Gln Thr Leu Arg Leu Gln Met Asn Phe Ile			
390	395	400	
aac cag gcc cag ctc ggc atc ttc agg gcc ttc cct ggc ctg cgc tac			1363
Asn Gln Ala Gln Leu Gly Ile Phe Arg Ala Phe Pro Gly Leu Arg Tyr			
405	410	415	

gtg gac ctg tcg gac aac cgc atc agc gga gct tcg gag ctg aca gcc	1411
Val Asp Leu Ser Asp Asn Arg Ile Ser Gly Ala Ser Glu Leu Thr Ala	
420 425 430 435	
acc atg ggg gag gca gat gga ggg gag aag gtc tgg ctg cag cct ggg	1459
Thr Met Gly Glu Ala Asp Gly Gly Glu Lys Val Trp Leu Gln Pro Gly	
440 445 450	
gac ctt gct ccg gcc cca gtg gac act ccc agc tct gaa gac ttc agg	1507
Asp Leu Ala Pro Ala Pro Val Asp Thr Pro Ser Ser Glu Asp Phe Arg	
455 460 465	
ccc aac tgc agc acc ctc aac ttc acc ttg gat ctg tca cgg aac aac	1555
Pro Asn Cys Ser Thr Leu Asn Phe Thr Leu Asp Leu Ser Arg Asn Asn	
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Leu Val Thr Val Gln Pro Glu Met Phe Ala Gln Leu Ser His Leu Gln	
485 490 495	
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Cys Leu Arg Leu Ser His Asn Cys Ile Ser Gln Ala Val Asn Gly Ser	
500 505 510 515	
cag ttc ctg ccg ctg acc ggt ctg cag gtg cta gac ctg tcc cac aat	1699
Gln Phe Leu Pro Leu Thr Gly Leu Gln Val Leu Asp Leu Ser His Asn	
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aag ctg gac ctc tac cac gag cac tca ttc acg gag cta cca cga ctg	1747
Lys Leu Asp Leu Tyr His Glu His Ser Phe Thr Glu Leu Pro Arg Leu	
535 540 545	
gag gcc ctg gac ctc agc tac aac agc cag ccc ttt ggc atg cag ggc	1795
Glu Ala Leu Asp Leu Ser Tyr Asn Ser Gln Pro Phe Gly Met Gln Gly	
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gtg ggc cac aac ttc agc ttc gtg gct cac ctg cgc acc ctg cgc cac	1843
Val Gly His Asn Phe Ser Phe Val Ala His Leu Arg Thr Leu Arg His	
565 570 575	
ctc agc ctg gcc cac aac aac atc cac agc caa gtg tcc cag cag ctc	1891
Leu Ser Leu Ala His Asn Asn Ile His Ser Gln Val Ser Gln Gln Leu	
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tgc agt acg tcg ctg cgg gcc ctg gac ttc agc ggc aat gca ctg ggc	1939
Cys Ser Thr Ser Leu Arg Ala Leu Asp Phe Ser Gly Asn Ala Leu Gly	

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cat atg tgg gcc gag gga gac ctc tat ctg cac ttc ttc caa ggc ctg			1987
His Met Trp Ala Glu Gly Asp Leu Tyr Leu His Phe Phe Gln Gly Leu			
615	620	625	
agc ggt ttg atc tgg ctg gac ttg tcc cag aac cgc ctg cac acc ctc			2035
Ser Gly Leu Ile Trp Leu Asp Leu Ser Gln Asn Arg Leu His Thr Leu			
630	635	640	
ctg ccc caa acc ctg cgc aac ctc ccc aag agc cta cag gtg ctg cgt			2083
Leu Pro Gln Thr Leu Arg Asn Leu Pro Lys Ser Leu Gln Val Leu Arg			
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ctc cgt gac aat tac ctg gcc ttc ttt aag tgg tgg agc ctc cac ttc			2131
Leu Arg Asp Asn Tyr Leu Ala Phe Phe Lys Trp Trp Ser Leu His Phe			
660	665	670	675
ctg ccc aaa ctg gaa gtc ctc gac ctg gca gga aac cag ctg aag gcc			2179
Leu Pro Lys Leu Glu Val Leu Asp Leu Ala Gly Asn Gln Leu Lys Ala			
680	685	690	
ctg acc aat ggc agc ctg cct gct ggc acc cgg ctc cgg agg ctg gat			2227
Leu Thr Asn Gly Ser Leu Pro Ala Gly Thr Arg Leu Arg Arg Leu Asp			
695	700	705	
gtc agc tgc aac agc atc agc ttc gtg gcc ccc ggc ttc ttt tcc aag			2275
Val Ser Cys Asn Ser Ile Ser Phe Val Ala Pro Gly Phe Phe Ser Lys			
710	715	720	
gcc aag gag ctg cga gag ctc aac ctt agc gcc aac gcc ctc aag aca			2323
Ala Lys Glu Leu Arg Glu Leu Asn Leu Ser Ala Asn Ala Leu Lys Thr			
725	730	735	
gtg gac cac tcc tgg ttt ggg ccc ctg gcg agt gcc ctg caa ata cta			2371
Val Asp His Ser Trp Phe Gly Pro Leu Ala Ser Ala Leu Gln Ile Leu			
740	745	750	755
gat gta agc gcc aac cct ctg cac tgc gcc tgt ggg gcg gcc ttt atg			2419
Asp Val Ser Ala Asn Pro Leu His Cys Ala Cys Gly Ala Ala Phe Met			
760	765	770	
gac ttc ctg ctg gag gtg cag gct gcc gtg ccc ggt ctg ccc agc cgg			2467
Asp Phe Leu Leu Glu Val Gln Ala Ala Val Pro Gly Leu Pro Ser Arg			
775	780	785	

gtg aag tgt ggc agt ccg ggc cag ctc cag ggc ctc agc atc ttt gca	2515
Val Lys Cys Gly Ser Pro Gly Gln Leu Gln Gly Leu Ser Ile Phe Ala	
790 795 800	
cag gac ctg cgc ctc tgc ctg gat gag gcc ctc tcc tgg gac tgt ttc	2563
Gln Asp Leu Arg Leu Cys Leu Asp Glu Ala Leu Ser Trp Asp Cys Phe	
805 810 815	
gcc ctc tcg ctg ctg gct gtg gct ctg ggc ctg ggt gtg ccc atg ctg	2611
Ala Leu Ser Leu Leu Ala Val Ala Leu Gly Leu Gly Val Pro Met Leu	
820 825 830 835	
cat cac ctc tgt ggc tgg gac ctc tgg tac tgc ttc cac ctg tgc ctg	2659
His His Leu Cys Gly Trp Asp Leu Trp Tyr Cys Phe His Leu Cys Leu	
840 845 850	
gcc tgg ctt ccc tgg cgg ggg cgg caa agt ggg cga gat gag gat gcc	2707
Ala Trp Leu Pro Trp Arg Gly Arg Gln Ser Gly Arg Asp Glu Asp Ala	
855 860 865	
ctg ccc tac gat gcc ttc gtg gtc ttc gac aaa acg cag agc gca gtg	2755
Leu Pro Tyr Asp Ala Phe Val Val Phe Asp Lys Thr Gln Ser Ala Val	
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Ala Asp Trp Val Tyr Asn Glu Leu Arg Gly Gln Leu Glu Glu Cys Arg	
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Gly Arg Trp Ala Leu Arg Leu Cys Leu Glu Glu Arg Asp Trp Leu Pro	
900 905 910 915	
ggc aaa acc ctc ttt gag aac ctg tgg gcc tgc gtc tat ggc agc cgc	2899
Gly Lys Thr Leu Phe Glu Asn Leu Trp Ala Ser Val Tyr Gly Ser Arg	
920 925 930	
aag acg ctg ttt gtg ctg gcc cac acg gac cgg gtc agt ggt ctc ttg	2947
Lys Thr Leu Phe Val Leu Ala His Thr Asp Arg Val Ser Gly Leu Leu	
935 940 945	
cgc gcc agc ttc ctg ctg gcc cag cag cgc ctg ctg gag gac cgc aag	2995
Arg Ala Ser Phe Leu Leu Ala Gln Gln Arg Leu Leu Glu Asp Arg Lys	
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Asp Val Val Val Leu Val Ile Leu Ser Pro Asp Gly Arg Arg Ser Arg	

965	970	975	
tac gtg cgg ctg cgc cag cgc ctc tgc cgc cag agt gtc ctc ctc tgg			3091
Tyr Val Arg Leu Arg Gln Arg Leu Cys Arg Gln Ser Val Leu Leu Trp			
980	985	990	995
ccc cac cag ccc agt ggt cag cgc agc ttc tgg gcc cag ctg ggc atg			3139
Pro His Gln Pro Ser Gly Gln Arg Ser Phe Trp Ala Gln Leu Gly Met			
1000	1005	1010	
gcc ctg acc agg gac aac cac cac ttc tat aac cgg aac ttc tgc cag			3187
Ala Leu Thr Arg Asp Asn His His Phe Tyr Asn Arg Asn Phe Cys Gln			
1015	1020	1025	
gga ccc acg gcc gaa tag ccgtgagccg gaatcctgca cgggtgccacc			3235
Gly Pro Thr Ala Glu			
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 35 40 45
 Phe Leu Lys Ser Val Pro His Phe Ser Met Ala Ala Pro Arg Gly Asn
 50 55 60
 Val Thr Ser Leu Ser Leu Ser Ser Asn Arg Ile His His Leu His Asp
 65 70 75 80
 Ser Asp Phe Ala His Leu Pro Ser Leu Arg His Leu Asn Leu Lys Trp
 85 90 95
 Asn Cys Pro Pro Val Gly Leu Ser Pro Met His Phe Pro Cys His Met
 100 105 110
 Thr Ile Glu Pro Ser Thr Phe Leu Ala Val Pro Thr Leu Glu Glu Leu
 115 120 125
 Asn Leu Ser Tyr Asn Asn Ile Met Thr Val Pro Ala Leu Pro Lys Ser
 130 135 140
 Leu Ile Ser Leu Ser Leu Ser His Thr Asn Ile Leu Met Leu Asp Ser

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Ala	Ser	Leu	Ala	Gly	Leu	His	Ala	Leu	Arg	Phe	Leu	Phe	Met	Asp Gly
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Asn	Cys	Tyr	Tyr	Lys	Asn	Pro	Cys	Arg	Gln	Ala	Leu	Glu	Val	Ala Pro
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Gly	Ala	Leu	Leu	Gly	Leu	Gly	Asn	Leu	Thr	His	Leu	Ser	Leu	Lys Tyr
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Ala	Asn	Leu	Thr	Ala	Leu	Arg	Val	Leu	Asp	Val	Gly	Gly	Asn	Cys Arg
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Arg	Cys	Asp	His	Ala	Pro	Asn	Pro	Cys	Met	Glu	Cys	Pro	Arg	His Phe
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Pro	Gln	Leu	His	Pro	Asp	Thr	Phe	Ser	His	Leu	Ser	Arg	Leu	Glu Gly
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Leu	Val	Leu	Lys	Asp	Ser	Ser	Leu	Ser	Trp	Leu	Asn	Ala	Ser	Trp Phe
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Arg	Gly	Leu	Gly	Asn	Leu	Arg	Val	Leu	Asp	Leu	Ser	Glu	Asn	Phe Leu
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Tyr	Lys	Cys	Ile	Thr	Lys	Thr	Lys	Ala	Phe	Gln	Gly	Leu	Thr	Gln Leu
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Arg	Lys	Leu	Asn	Leu	Ser	Phe	Asn	Tyr	Gln	Lys	Arg	Val	Ser	Phe Ala
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His	Leu	Ser	Leu	Ala	Pro	Ser	Phe	Gly	Ser	Leu	Val	Ala	Leu	Lys Glu
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Leu	Asp	Met	His	Gly	Ile	Phe	Phe	Arg	Ser	Leu	Asp	Glu	Thr	Thr Leu
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Arg	Pro	Leu	Ala	Arg	Leu	Pro	Met	Leu	Gln	Thr	Leu	Arg	Leu	Gln Met
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Asn	Phe	Ile	Asn	Gln	Ala	Gln	Leu	Gly	Ile	Phe	Arg	Ala	Phe	Pro Gly
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Leu	Arg	Tyr	Val	Asp	Leu	Ser	Asp	Asn	Arg	Ile	Ser	Gly	Ala	Ser Glu
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Leu	Thr	Ala	Thr	Met	Gly	Glu	Ala	Asp	Gly	Gly	Glu	Lys	Val	Trp Leu
	435						440					445		
Gln	Pro	Gly	Asp	Leu	Ala	Pro	Ala	Pro	Val	Asp	Thr	Pro	Ser	Ser Glu
	450					455				460				
Asp	Phe	Arg	Pro	Asn	Cys	Ser	Thr	Leu	Asn	Phe	Thr	Leu	Asp	Leu Ser
465				470					475					480
Arg	Asn	Asn	Leu	Val	Thr	Val	Gln	Pro	Glu	Met	Phe	Ala	Gln	Leu Ser
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His	Leu	Gln	Cys	Leu	Arg	Leu	Ser	His	Asn	Cys	Ile	Ser	Gln	Ala Val
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Asn	Gly	Ser	Gln	Phe	Leu	Pro	Leu	Thr	Gly	Leu	Gln	Val	Leu	Asp Leu

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Ser His Asn Lys Leu Asp	Leu Tyr His Glu His	Ser Phe Thr Glu Leu
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Pro Arg Leu Glu Ala Leu	Asp Leu Ser Tyr Asn	Ser Gln Pro Phe Gly
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Met Gln Gly Val Gly His	Asn Phe Ser Phe Val	Ala His Leu Arg Thr
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Leu Arg His Leu Ser Leu	Ala His Asn Asn Ile	His Ser Gln Val Ser
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Gln Gln Leu Cys Ser Thr	Ser Leu Arg Ala Leu	Asp Phe Ser Gly Asn
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Ala Leu Gly His Met Trp	Ala Glu Gly Asp Leu	Tyr Leu His Phe Phe
610	615	620
Gln Gly Leu Ser Gly Leu	Ile Trp Leu Asp Leu	Ser Gln Asn Arg Leu
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His Thr Leu Leu Pro Gln	Thr Leu Arg Asn Leu	Pro Lys Ser Leu Gln
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Val Leu Arg Leu Arg Asp	Asn Tyr Leu Ala Phe	Phe Lys Trp Trp Ser
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675	680	685
Leu Lys Ala Leu Thr Asn	Gly Ser Leu Pro Ala	Gly Thr Arg Leu Arg
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Arg Leu Asp Val Ser Cys	Asn Ser Ile Ser Phe	Val Ala Pro Gly Phe
705	710	715
Phe Ser Lys Ala Lys Glu	Leu Arg Glu Leu Asn	Leu Ser Ala Asn Ala
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Leu Lys Thr Val Asp His	Ser Trp Phe Gly Pro	Leu Ala Ser Ala Leu
740	745	750
Gln Ile Leu Asp Val Ser	Ala Asn Pro Leu His	Cys Ala Cys Gly Ala
755	760	765
Ala Phe Met Asp Phe Leu	Leu Glu Val Gln Ala	Ala Val Pro Gly Leu
770	775	780
Pro Ser Arg Val Lys Cys	Gly Ser Pro Gly Gln	Leu Gln Gly Leu Ser
785	790	795
Ile Phe Ala Gln Asp Leu	Arg Leu Cys Leu Asp	Glu Ala Leu Ser Trp
805	810	815
Asp Cys Phe Ala Leu Ser	Leu Leu Ala Val Ala	Leu Gly Leu Gly Val
820	825	830
Pro Met Leu His His Leu	Cys Gly Trp Asp Leu	Trp Tyr Cys Phe His
835	840	845
Leu Cys Leu Ala Trp Leu	Pro Trp Arg Gly Arg	Gln Ser Gly Arg Asp
850	855	860
Glu Asp Ala Leu Pro Tyr	Asp Ala Phe Val Val	Phe Asp Lys Thr Gln
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Ser Ala Val Ala Asp Trp	Val Tyr Asn Glu Leu	Arg Gly Gln Leu Glu

885							890							895			
Glu	Cys	Arg	Gly	Arg	Trp	Ala	Leu	Arg	Leu	Cys	Leu	Glu	Glu	Arg	Asp		
900							905							910			
Trp	Leu	Pro	Gly	Lys	Thr	Leu	Phe	Glu	Asn	Leu	Trp	Ala	Ser	Val	Tyr		
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Gly	Ser	Arg	Lys	Thr	Leu	Phe	Val	Leu	Ala	His	Thr	Asp	Arg	Val	Ser		
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Gly	Leu	Leu	Arg	Ala	Ser	Phe	Leu	Leu	Ala	Gln	Gln	Arg	Leu	Leu	Glu		
945							950							955			
Asp	Arg	Lys	Asp	Val	Val	Val	Leu	Val	Ile	Leu	Ser	Pro	Asp	Gly	Arg		
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Arg	Ser	Arg	Tyr	Val	Arg	Leu	Arg	Gln	Arg	Leu	Cys	Arg	Gln	Ser	Val		
980							985							990			
Leu	Leu	Trp	Pro	His	Gln	Pro	Ser	Gly	Gln	Arg	Ser	Phe	Trp	Ala	Gln		
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Leu	Gly	Met	Ala	Leu	Thr	Arg	Asp	Asn	His	His	Phe	Tyr	Asn	Arg	Asn		
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Met Val Leu

1

cgt cga agg act ctg cac ccc ttg tcc clc ctg gta cag gcc gca gtg 163
Arg Arg Arg Thr Leu His Pro Leu Ser Leu Leu Val Gln Ala Ala Val

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Leu Ala Glu Thr Leu Ala Leu Gly Thr Leu Pro Ala Phe Leu Pro Cys
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Glu Leu Lys Pro His Gly Leu Val Asp Cys Asn Trp Leu Phe Leu Lys	
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Ser Val Pro Arg Phe Ser Ala Ala Ala Ser Cys Ser Asn Ile Thr Arg	
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Leu Ser Leu Ile Ser Asn Arg Ile His His Leu His Asn Ser Asp Phe	
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Val His Leu Ser Asn Leu Arg Gln Leu Asn Leu Lys Trp Asn Cys Pro	
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Pro Thr Gly Leu Ser Pro Leu His Phe Ser Cys His Met Thr Ile Glu	
100 105 110 115	
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Pro Arg Thr Phe Leu Ala Met Arg Thr Leu Glu Glu Leu Asn Leu Ser	
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tat aat ggt atc acc act gtg ccc cga ctg ccc agc tcc ctg gtg aat	547
Tyr Asn Gly Ile Thr Thr Val Pro Arg Leu Pro Ser Ser Leu Val Asn	
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Leu Ser Leu Ser His Thr Asn Ile Leu Val Leu Asp Ala Asn Ser Leu	
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gcc ggc cta tac agc ctg cgc gtt ctc ttc atg gac ggg aac tgc tac	643
Ala Gly Leu Tyr Ser Leu Arg Val Leu Phe Met Asp Gly Asn Cys Tyr	
165 170 175	
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Tyr Lys Asn Pro Cys Thr Gly Ala Val Lys Val Thr Pro Gly Ala Leu	
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Leu Gly Leu Ser Asn Leu Thr His Leu Ser Val Lys Tyr Asn Asn Leu	
200 205 210	
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Thr Lys Val Pro Arg Gln Leu Pro Pro Ser Leu Glu Tyr Leu Leu Val	

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Ser Tyr Asn Leu Ile Val Lys Leu Gly Pro Glu Asp Leu Ala Asn Leu			
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Thr Ser Leu Arg Val Leu Asp Val Gly Gly Asn Cys Arg Arg Cys Asp			
245	250	255	
cat gcc ccc aat ccc tgt ata gaa tgt ggc caa aag tcc ctc cac ctg			931
His Ala Pro Asn Pro Cys Ile Glu Cys Gly Gln Lys Ser Leu His Leu			
260	265	270	275
cac cct gag acc ttc cat cac ctg agc cat ctg gaa ggc ctg gig ctg			979
His Pro Glu Thr Phe His His Leu Ser His Leu Glu Gly Leu Val Leu			
280	285	290	
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Lys Asp Ser Ser Leu His Thr Leu Asn Ser Ser Trp Phe Gln Gly Leu			
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Val Asn Leu Ser Val Leu Asp Leu Ser Glu Asn Phe Leu Tyr Glu Ser			
310	315	320	
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Ile Asn His Thr Asn Ala Phe Gln Asn Leu Thr Arg Leu Arg Lys Leu			
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aac ctg tcc ttc aat tac cgc aag aag gta tcc ttt gcc cgc ctc cac			1171
Asn Leu Ser Phe Asn Tyr Arg Lys Lys Val Ser Phe Ala Arg Leu His			
340	345	350	355
ctg gca agt tcc ttc aag aac ctg gig tca ctg cag gag ctg aac atg			1219
Leu Ala Ser Ser Phe Lys Asn Leu Val Ser Leu Gln Glu Leu Asn Met			
360	365	370	
aac ggc atc ttc ttc cgc tcg ctc aac aag tac acg ctc aga tgg ctg			1267
Asn Gly Ile Phe Phe Arg Ser Leu Asn Lys Tyr Thr Leu Arg Trp Leu			
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Ala Asp Leu Pro Lys Leu His Thr Leu His Leu Gln Met Asn Phe Ile			
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Val Asp Leu Ser Asp Asn Arg Ile Ser Gly Pro Ser Thr Leu Ser Glu	
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Ala Asp Pro His Pro Ala Pro Leu Ser Thr Pro Ala Ser Lys Asn Phe	
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Met Asp Arg Cys Lys Asn Phe Lys Phe Thr Met Asp Leu Ser Arg Asn	
470 475 480	
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Asn Leu Val Thr Ile Lys Pro Glu Met Phe Val Asn Leu Ser Arg Leu	
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Gln Cys Leu Ser Leu Ser His Asn Ser Ile Ala Gln Ala Val Asn Gly	
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tct cag ttc ctg ccg ctg act aat ctg cag gtg ctg gac ctg tcc cat	1699
Ser Gln Phe Leu Pro Leu Thr Asn Leu Gln Val Leu Asp Leu Ser His	
520 525 530	
aac aaa ctg gac ttg tac cac tgg aaa tcg ttc agt gag cta cca cag	1747
Asn Lys Leu Asp Leu Tyr His Trp Lys Ser Phe Ser Glu Leu Pro Gln	
535 540 545	
ttg cag gcc ctg gac ctg agc tac aac agc cag ccc ttt agc atg aag	1795
Leu Gln Ala Leu Asp Leu Ser Tyr Asn Ser Gln Pro Phe Ser Met Lys	
550 555 560	
ggt ata ggc cac aat ttc agt ttt gtg gcc cat ctg tcc atg cta cac	1843
Gly Ile Gly His Asn Phe Ser Phe Val Ala His Leu Ser Met Leu His	
565 570 575	
agc ctt agc ctg gca cac aat gac att cat acc cgt gtg tcc tca cat	1891
Ser Leu Ser Leu Ala His Asn Asp Ile His Thr Arg Val Ser Ser His	

580	585	590	595	
ctc aac agc aac tca gtg agg ttt ctt gac ttc agc ggc aac ggt atg				1939
Leu Asn Ser Asn Ser Val Arg Phe Leu Asp Phe Ser Gly Asn Gly Met				
	600	605	610	
ggc cgc atg tgg gat gag ggg ggc ctt tat ctc cat ttc ttc caa ggc				1987
Gly Arg Met Trp Asp Glu Gly Gly Leu Tyr Leu His Phe Phe Gln Gly				
	615	620	625	
ctg agt ggc ctg ctg aag ctg gac ctg tct caa aat aac ctg cat atc				2035
Leu Ser Gly Leu Leu Lys Leu Asp Leu Ser Gln Asn Asn Leu His Ile				
	630	635	640	
ctc cgg ccc cag aac ctt gac aac ctc ccc aag agc ctg aag ctg ctg				2083
Leu Arg Pro Gln Asn Leu Asp Asn Leu Pro Lys Ser Leu Lys Leu Leu				
	645	650	655	
agc ctc cga gac aac tac cta tct ttc ttt aac tgg acc agt ctg tcc				2131
Ser Leu Arg Asp Asn Tyr Leu Ser Phe Phe Asn Trp Thr Ser Leu Ser				
660	665	670	675	
ttc ctg ccc aac ctg gaa gtc cta gac ctg gca ggc aac cag cta aag				2179
Phe Leu Pro Asn Leu Glu Val Leu Asp Leu Ala Gly Asn Gln Leu Lys				
	680	685	690	
gcc ctg acc aat ggc acc ctg cct aat ggc acc ctc ctc cag aaa ctg				2227
Ala Leu Thr Asn Gly Thr Leu Pro Asn Gly Thr Leu Leu Gln Lys Leu				
	695	700	705	
gat gtc agc agc aac agt atc gtc tct gtg gtc cca gcc ttc ttc gct				2275
Asp Val Ser Ser Asn Ser Ile Val Ser Val Val Pro Ala Phe Phe Ala				
	710	715	720	
ctg gcg gtc gag ctg aaa gag gtc aac ctc agc cac aac att ctc aag				2323
Leu Ala Val Glu Leu Lys Glu Val Asn Leu Ser His Asn Ile Leu Lys				
	725	730	735	
acg gtg gat cgc tcc tgg ttt ggg ccc att gtg atg aac ctg aca gtt				2371
Thr Val Asp Arg Ser Trp Phe Gly Pro Ile Val Met Asn Leu Thr Val				
740	745	750	755	
cta gac gtg aga agc aac cct ctg cac tgt gcc tgt ggg gca gcc ttc				2419
Leu Asp Val Arg Ser Asn Pro Leu His Cys Ala Cys Gly Ala Ala Phe				
	760	765	770	

gta gac tta ctg ttg gag gtg cag acc aag gtg cct ggc ctg gct aat	2467
Val Asp Leu Leu Leu Glu Val Gln Thr Lys Val Pro Gly Leu Ala Asn	
775 780 785	
ggt gtg aag tgt ggc agc ccc ggc cag ctg cag ggc cgt agc atc ttc	2515
Gly Val Lys Cys Gly Ser Pro Gly Gln Leu Gln Gly Arg Ser Ile Phe	
790 795 800	
gca cag gac ctg cgg ctg tgc ctg gat gag gtc ctc tct tgg gac tgc	2563
Ala Gln Asp Leu Arg Leu Cys Leu Asp Glu Val Leu Ser Trp Asp Cys	
805 810 815	
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Phe Gly Leu Ser Leu Leu Ala Val Ala Val Gly Met Val Val Pro Ile	
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Leu His His Leu Cys Gly Trp Asp Val Trp Tyr Cys Phe His Leu Cys	
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Leu Pro Tyr Asp Ala Phe Val Val Phe Asp Lys Ala Gln Ser Ala Val	
870 875 880	
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Ala Asp Trp Val Tyr Asn Glu Leu Arg Val Arg Leu Glu Glu Arg Arg	
885 890 895	
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Gly Arg Arg Ala Leu Arg Leu Cys Leu Glu Asp Arg Asp Trp Leu Pro	
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Lys Thr Leu Phe Val Leu Ala His Thr Asp Arg Val Ser Gly Leu Leu	
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cgc acc agc ttc ctg ctg gct cag cag cgc ctg ttg gaa gac cgc aag	2995
Arg Thr Ser Phe Leu Leu Ala Gln Gln Arg Leu Leu Glu Asp Arg Lys	

950 955 960
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 Asp Val Val Val Leu Val Ile Leu Arg Pro Asp Ala His Arg Ser Arg
 965 970 975
 tai gtg cga ctg cgc cag cgt ctc tgc cgc cag agt gtg ctc ttc tgg 3091
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 Ala Leu Thr Arg Asp Asn Arg His Phe Tyr Asn Gln Asn Phe Cys Arg
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 20 25 30
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 35 40 45
 Phe Leu Lys Ser Val Pro Arg Phe Ser Ala Ala Ala Ser Cys Ser Asn
 50 55 60
 Ile Thr Arg Leu Ser Leu Ile Ser Asn Arg Ile His His Leu His Asn

65					70					75				80
Ser	Asp	Phe	Val	His	Leu	Ser	Asn	Leu	Arg	Gln	Leu	Asn	Leu	Lys
				85					90					95
Asn	Cys	Pro	Pro	Thr	Gly	Leu	Ser	Pro	Leu	His	Phe	Ser	Cys	His
		100						105					110	Met
Thr	Ile	Glu	Pro	Arg	Thr	Phe	Leu	Ala	Met	Arg	Thr	Leu	Glu	Glu
		115					120					125		Leu
Asn	Leu	Ser	Tyr	Asn	Gly	Ile	Thr	Thr	Val	Pro	Arg	Leu	Pro	Ser
	130					135				140				Ser
Leu	Val	Asn	Leu	Ser	Leu	Ser	His	Thr	Asn	Ile	Leu	Val	Leu	Asp
145					150				155					Ala
Asn	Ser	Leu	Ala	Gly	Leu	Tyr	Ser	Leu	Arg	Val	Leu	Phe	Met	Asp
				165					170				175	Gly
Asn	Cys	Tyr	Tyr	Lys	Asn	Pro	Cys	Thr	Gly	Ala	Val	Lys	Val	Thr
		180						185					190	Pro
Gly	Ala	Leu	Leu	Gly	Leu	Ser	Asn	Leu	Thr	His	Leu	Ser	Val	Lys
	195						200					205		Tyr
Asn	Asn	Leu	Thr	Lys	Val	Pro	Arg	Gln	Leu	Pro	Pro	Ser	Leu	Glu
	210					215				220				Tyr
Leu	Leu	Val	Ser	Tyr	Asn	Leu	Ile	Val	Lys	Leu	Gly	Pro	Glu	Asp
225					230					235				Leu
Ala	Asn	Leu	Thr	Ser	Leu	Arg	Val	Leu	Asp	Val	Gly	Gly	Asn	Cys
				245					250				255	Arg
Arg	Cys	Asp	His	Ala	Pro	Asn	Pro	Cys	Ile	Glu	Cys	Gly	Gln	Lys
		260						265					270	Ser
Leu	His	Leu	His	Pro	Glu	Thr	Phe	His	His	Leu	Ser	His	Leu	Glu
	275					280						285		Gly
Leu	Val	Leu	Lys	Asp	Ser	Ser	Leu	His	Thr	Leu	Asn	Ser	Ser	Trp
	290				295						300			Phe
Gln	Gly	Leu	Val	Asn	Leu	Ser	Val	Leu	Asp	Leu	Ser	Glu	Asn	Phe
305				310					315					Leu
Tyr	Glu	Ser	Ile	Asn	His	Thr	Asn	Ala	Phe	Gln	Asn	Leu	Thr	Arg
			325					330					335	Leu
Arg	Lys	Leu	Asn	Leu	Ser	Phe	Asn	Tyr	Arg	Lys	Lys	Val	Ser	Phe
		340					345					350		Ala
Arg	Leu	His	Leu	Ala	Ser	Ser	Phe	Lys	Asn	Leu	Val	Ser	Leu	Gln
	355					360					365			Glu
Leu	Asn	Met	Asn	Gly	Ile	Phe	Phe	Arg	Ser	Leu	Asn	Lys	Tyr	Thr
	370				375					380				Leu
Arg	Trp	Leu	Ala	Asp	Leu	Pro	Lys	Leu	His	Thr	Leu	His	Leu	Gln
385				390					395					Met
Asn	Phe	Ile	Asn	Gln	Ala	Gln	Leu	Ser	Ile	Phe	Gly	Thr	Phe	Arg
			405					410					415	Ala
Leu	Arg	Phe	Val	Asp	Leu	Ser	Asp	Asn	Arg	Ile	Ser	Gly	Pro	Ser
		420					425					430		Thr
Leu	Ser	Glu	Ala	Thr	Pro	Glu	Glu	Ala	Asp	Asp	Ala	Glu	Gln	Glu

435		440		445
Leu Leu Ser Ala Asp Pro His Pro Ala Pro Leu Ser Thr Pro Ala Ser				
450		455		460
Lys Asn Phe Met Asp Arg Cys Lys Asn Phe Lys Phe Thr Met Asp Leu				
465		470		475
Ser Arg Asn Asn Leu Val Thr Ile Lys Pro Glu Met Phe Val Asn Leu				480
	485		490	495
Ser Arg Leu Gln Cys Leu Ser Leu Ser His Asn Ser Ile Ala Gln Ala				
	500		505	510
Val Asn Gly Ser Gln Phe Leu Pro Leu Thr Asn Leu Gln Val Leu Asp				
	515		520	525
Leu Ser His Asn Lys Leu Asp Leu Tyr His Trp Lys Ser Phe Ser Glu				
	530		535	540
Leu Pro Gln Leu Gln Ala Leu Asp Leu Ser Tyr Asn Ser Gln Pro Phe				
545		550		555
Ser Met Lys Gly Ile Gly His Asn Phe Ser Phe Val Ala His Leu Ser				560
	565		570	575
Met Leu His Ser Leu Ser Leu Ala His Asn Asp Ile His Thr Arg Val				
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Ser Ser His Leu Asn Ser Asn Ser Val Arg Phe Leu Asp Phe Ser Gly				
	595		600	605
Asn Gly Met Gly Arg Met Trp Asp Glu Gly Gly Leu Tyr Leu His Phe				
	610		615	620
Phe Gln Gly Leu Ser Gly Leu Leu Lys Leu Asp Leu Ser Gln Asn Asn				
625		630		635
Leu His Ile Leu Arg Pro Gln Asn Leu Asp Asn Leu Pro Lys Ser Leu				640
	645		650	655
Lys Leu Leu Ser Leu Arg Asp Asn Tyr Leu Ser Phe Phe Asn Trp Thr				
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Ser Leu Ser Phe Leu Pro Asn Leu Glu Val Leu Asp Leu Ala Gly Asn				
	675		680	685
Gln Leu Lys Ala Leu Thr Asn Gly Thr Leu Pro Asn Gly Thr Leu Leu				
	690		695	700
Gln Lys Leu Asp Val Ser Ser Asn Ser Ile Val Ser Val Val Pro Ala				
705		710		715
Phe Phe Ala Leu Ala Val Glu Leu Lys Glu Val Asn Leu Ser His Asn				720
	725		730	735
Ile Leu Lys Thr Val Asp Arg Ser Trp Phe Gly Pro Ile Val Met Asn				
	740		745	750
Leu Thr Val Leu Asp Val Arg Ser Asn Pro Leu His Cys Ala Cys Gly				
	755		760	765
Ala Ala Phe Val Asp Leu Leu Leu Glu Val Gln Thr Lys Val Pro Gly				
	770		775	780
Leu Ala Asn Gly Val Lys Cys Gly Ser Pro Gly Gln Leu Gln Gly Arg				
785		790		795
Ser Ile Phe Ala Gln Asp Leu Arg Leu Cys Leu Asp Glu Val Leu Ser				800

			805				810				815					
Trp	Asp	Cys	Phe	Gly	Leu	Ser	Leu	Leu	Ala	Val	Ala	Val	Gly	Met	Val	
			820				825				830					
Val	Pro	Ile	Leu	His	His	Leu	Cys	Gly	Trp	Asp	Val	Trp	Tyr	Cys	Phe	
			835				840				845					
His	Leu	Cys	Leu	Ala	Trp	Leu	Pro	Leu	Leu	Ala	Arg	Ser	Arg	Arg	Ser	
			850				855				860					
Ala	Gln	Ala	Leu	Pro	Tyr	Asp	Ala	Phe	Val	Val	Phe	Asp	Lys	Ala	Gln	
			865				870				875				880	
Ser	Ala	Val	Ala	Asp	Trp	Val	Tyr	Asn	Glu	Leu	Arg	Val	Arg	Leu	Glu	
			885				890				895					
Glu	Arg	Arg	Gly	Arg	Arg	Ala	Leu	Arg	Leu	Cys	Leu	Glu	Asp	Arg	Asp	
			900				905				910					
Trp	Leu	Pro	Gly	Gln	Thr	Leu	Phe	Glu	Asn	Leu	Trp	Ala	Ser	Ile	Tyr	
			915				920				925					
Gly	Ser	Arg	Lys	Thr	Leu	Phe	Val	Leu	Ala	His	Thr	Asp	Arg	Val	Ser	
			930				935				940					
Gly	Leu	Leu	Arg	Thr	Ser	Phe	Leu	Leu	Ala	Gln	Gln	Arg	Leu	Leu	Glu	
			945				950				955				960	
Asp	Arg	Lys	Asp	Val	Val	Val	Leu	Val	Ile	Leu	Arg	Pro	Asp	Ala	His	
			965				970				975					
Arg	Ser	Arg	Tyr	Val	Arg	Leu	Arg	Gln	Arg	Leu	Cys	Arg	Gln	Ser	Val	
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Leu	Phe	Trp	Pro	Gln	Gln	Pro	Asn	Gly	Gln	Gly	Gly	Phe	Trp	Ala	Gln	
			995				1000				1005					
Leu	Ser	Thr	Ala	Leu	Thr	Arg	Asp	Asn	Arg	His	Phe	Tyr	Asn	Gln	Asn	
			1010				1015				1020					
Phe	Cys	Arg	Gly	Pro	Thr	Ala	Glu									
			1025				1030									

<210> 5

$\langle 211 \rangle$ 20

<212> DNA

〈213〉 Artificial Sequence

 $\langle 220 \rangle$

<223> Description of Artificial Sequence:CpG ODN

<400> 5

tccatgacgt tcctgatgct

20

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CLAIMS:

1. A process for generating a homozygote non-human animal whose gene function encoding Toll-like Receptor (TLR) 9 protein is inactivated by a genetic mutation, which
5 process comprises:

(1) providing a vector containing a gene encoding TLR9 protein having the amino acid sequence shown in SEQ ID NO: 2;

(2) substituting whole or part of the gene
10 encoding TLR9 protein with pMC1 neo gene cassette and then introducing diphtheria toxin A fragment (DT-A) gene or herpes simplex virus thymidine kinase (HSV-tk) gene on a 3'-end side of the gene encoding TLR9 protein, to produce a targeting vector;

15 (3) linearizing and introducing the targeting vector into embryonic stem (ES) cells of the non-human animal, performing homologous recombination, and selecting ES cells which are targeted recombinants;

(4) introducing the selected ES cells into
20 blastocysts of the non-human animal;

(5) returning the blastocysts into a recipient non-human animal and producing a chimera non-human animal;

(6) intercrossing the produced chimera non-human animal with a wild-type non-human animal to produce
25 heterozygote non-human animals;

(7) intercrossing the heterozygote non-human animals to produce a homozygote non-human animal according to the Mendel's Law; and

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(8) confirming whether the produced homozygote non-human animal has its gene function encoding the TLR9 protein inactivated.

2. The process according to claim 1, wherein the gene
5 encoding TLR9 protein has the base sequence shown in SEQ ID NO: 1.

3. The process according to claim 1 or 2, wherein herpes simplex virus thymidine kinase (HSV-tk) gene is introduced in step (2).

10 4. The process according to any one of claims 1 to 3, wherein a male of the chimera non-human animal produced in step (5) is intercrossed with a female of the wild-type non-human animal in step (6).

5. The process according to any one of claims 1 to 4,
15 wherein the non-human animal is a rodent.

6. The process according to claim 5, wherein the rodent is a mouse.

7. An expression vector comprising:

(a) DNA encoding Toll-like Receptor (TLR) 9
20 protein having the amino acid sequence shown in SEQ ID NO: 2, and

(b) a control sequence which causes expression of TLR9 protein.

8. pBluescript II SK (+) vector containing therein
25 DNA encoding Toll-like Receptor (TLR) 9 protein having the amino acid sequence shown in SEQ ID NO: 2.

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9. The vector according to claim 7 or 8, wherein the DNA has the base sequence shown in SEQ ID NO: 1.

10. A host cell comprising an expression system which expresses Toll-like Receptor (TLR) 9 protein and comprises:

5 (a) DNA encoding TLR9 protein having the amino acid sequence shown in SEQ ID NO: 2, and

(b) a control sequence which causes expression of TLR protein.

11. The host cell according to claim 10, wherein the
10 DNA has the base sequence shown in SEQ ID NO: 1.

12. A fusion protein comprising:

(i) a protein comprising the amino acid sequence shown in SEQ ID NO: 2 or 4, and

(ii) a marker protein or a peptide tag, fused with
15 the protein (i).

13. The fusion protein according to claim 12, wherein the protein (i) comprises the amino acid sequence shown in SEQ ID NO: 2.

14. The fusion protein according to claim 12 or 13,
20 which comprises alkaline phosphatase, Fc region of an antibody, HRP or GFP, as the marker protein.

15. The fusion protein according to claim 12 or 13, which comprises Myc tag, His tag, FLAG tag or GST tag as the peptide tag.

25 16. An antibody which specifically binds to a protein comprising the amino acid sequence shown in SEQ ID NO: 2 or 4.

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17. The antibody according to claim 16, wherein the protein comprises the amino acid sequence shown in SEQ ID NO: 2.

18. A method for determining whether a target
5 substance is an agonist or an antagonist of Toll-like Receptor (TLR) 9 protein, which comprises:

culturing in vitro the host cell as defined in claim 10 or 11 in the presence of the target substance;

measuring TLR activity of the host cell; and

10 comparing the measured TLR activity with that of the host cell when cultured in the absence of the target substance.

19. The method according to claim 18, wherein the host cell is a bacteria cell, a fungus cell, an insect cell or an
15 animal or plant cell.

20. A kit for detecting a mutation in a gene encoding Toll-like Receptor (TLR) 9 protein having the amino acid sequence shown in SEQ ID NO: 2, comprising:

(a) a DNA having the base sequence shown in SEQ ID
20 NO: 1, and

(b) a reagent for performing PCR of a sequence of DNA encoding the TLR9 protein of a cell from a test subject.

21. A process for generating a cell from a homozygote non-human animal whose gene function encoding Toll-like
25 Receptor (TLR) 9 protein is inactivated by a genetic mutation, which process comprises:

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(1) providing a vector containing a gene encoding TLR9 protein having the amino acid sequence shown in SEQ ID NO: 2;

(2) substituting whole or part of the gene
5 encoding TLR9 protein with pMC1 neo gene cassette and then introducing diphtheria toxin A fragment (DT-A) gene or herpes simplex virus thymidine kinase (HSV-tk) gene on a 3'-end side of the gene encoding TLR9 protein, to produce a targeting vector;

10 (3) linearizing and introducing the targeting vector into embryonic stem (ES) cells of the non-human animal, performing homologous recombination, and selecting ES cells which are targeted recombinants;

(4) introducing the selected ES cells into
15 blastocysts of the non-human animal;

(5) returning the blastocysts into a recipient non-human animal and producing a chimera non-human animal;

(6) intercrossing the produced chimera non-human animal with a wild-type non-human animal to produce
20 heterozygote non-human animals;

(7) intercrossing the heterozygote non-human animals to produce a homozygote non-human animal according to the Mendel's Law;

(8) confirming whether the produced homozygote
25 non-human animal has its gene function encoding the TLR9 protein inactivated; and

(9) obtaining a cell from the homozygote non-human animal confirmed in step (8).

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22. The process according to claim 21, wherein the gene encoding TLR9 protein has the base sequence shown in SEQ ID NO: 1.

23. The process according to claim 21 or 22, wherein
5 herpes simplex virus thymidine kinase (HSV-tk) gene is introduced in step (2).

24. The process according to any one of claims 21 to 23, wherein a male of the chimera non-human animal produced in step (5) is intercrossed with a female of the wild-type
10 non-human animal in step (6).

25. The process according to any one of claims 21 to 24, wherein the non-human animal is a rodent.

26. The process according to claim 25, wherein the rodent is a mouse.

15 27. An isolated cell of a homozygote non-human animal, the cell having gene function encoding Toll-like Receptor (TLR) 9 protein having the amino acid sequence shown in SEQ ID NO: 2 inactivated by a genetic mutation and being obtained by the process as defined in any one of claims 21
20 to 26.

SMART & BIGGAR
OTTAWA, CANADA
PATENT AGENTS

FIG. 1

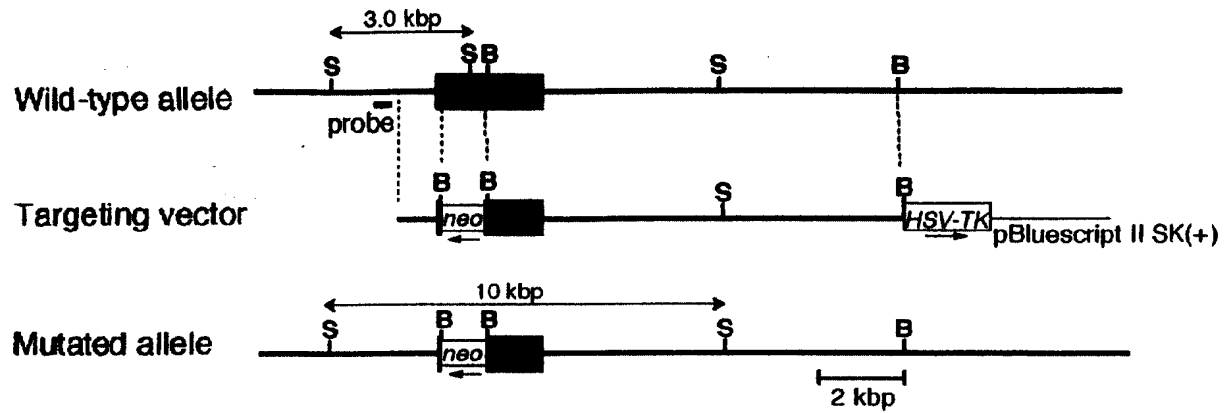


FIG. 2

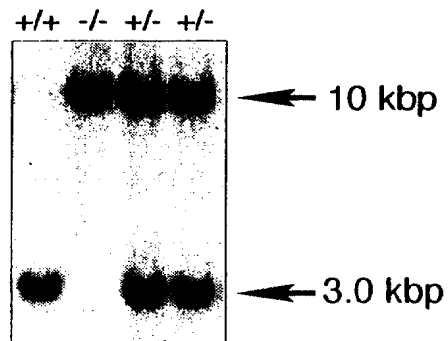


FIG. 3

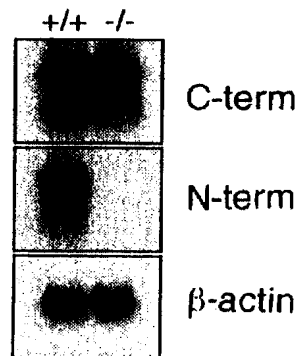


FIG. 4

+/+ :	87	90			96			100			110													
	TCC	AAC	CTG	CGG	CAG	CTG	AAC	CTC	AAG	TGG	AAC	TGT	CCA	CCC	ACT	GGC	CTT	AGC	CCC	TTG	CAC	TTC	TCT	TGC
	S	N	L	R	Q	L	N	L	K	W	N	C	P	P	T	G	L	S	P	L	H	F	S	C
-/- :	S	N	L	R	Q	L	N	L	K	W	I	L	S	T	C	P	R	R	I	R	T	N	D	P
	TCC	AAC	CTG	CGG	CAG	CTG	AAC	CTC	AAG	TGG	ATT	TTG	TCC	ACC	TGT	CCT	CGA	CGG	ATC	CGA	ACA	AAC	GAC	CCA
	87	90			96																			

+/+ :	120												130											
	CAC	ATG	ACC	ATT	GAG	CCC	AGA	ACC	TTC	CTG	GCT	ATG	CGT	ACA	CTG	GAG	GAG	CTG	AAC	CTG	AGC	TAT	AAT	GGT
	H	M	T	I	E	P	R	T	F	L	A	M	R	T	L	E	E	L	N	L	S	Y	N	G
-/- :	T	P	V	R	F	I	L	S	F	Y	C	R	S	P	Q	K	N	S	S	R	R	R	*	
	ACA	CCC	GTG	CGT	TTT	ATT	CTG	TCT	TTT	TAT	TGC	CGA	TCC	CCT	CAG	AAG	AAC	TGC	TCA	AGA	AGG	CGA	TAG	

FIG. 5

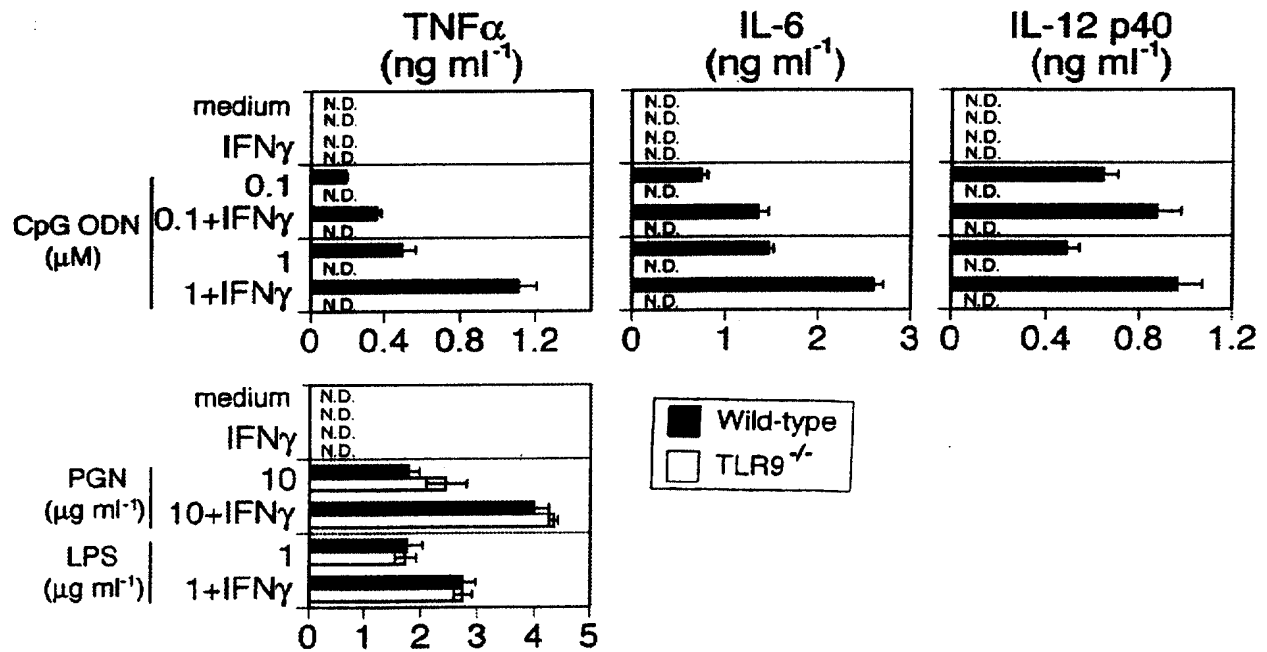


FIG. 6

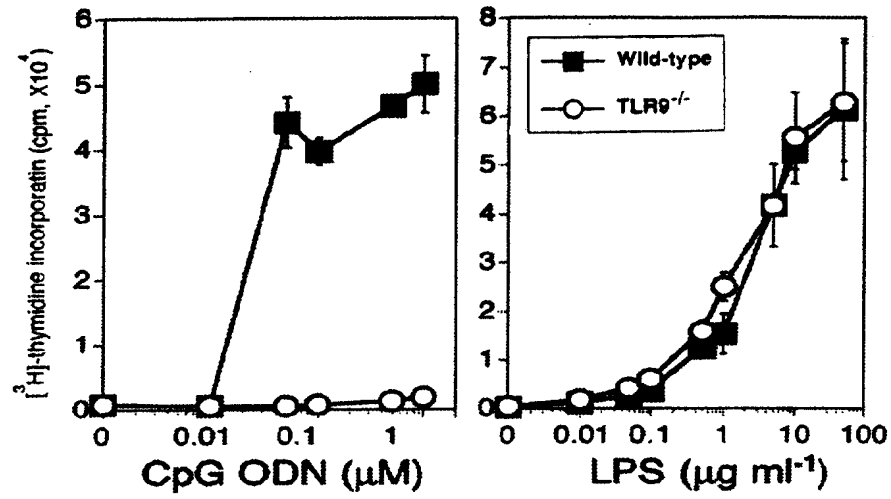


FIG. 7

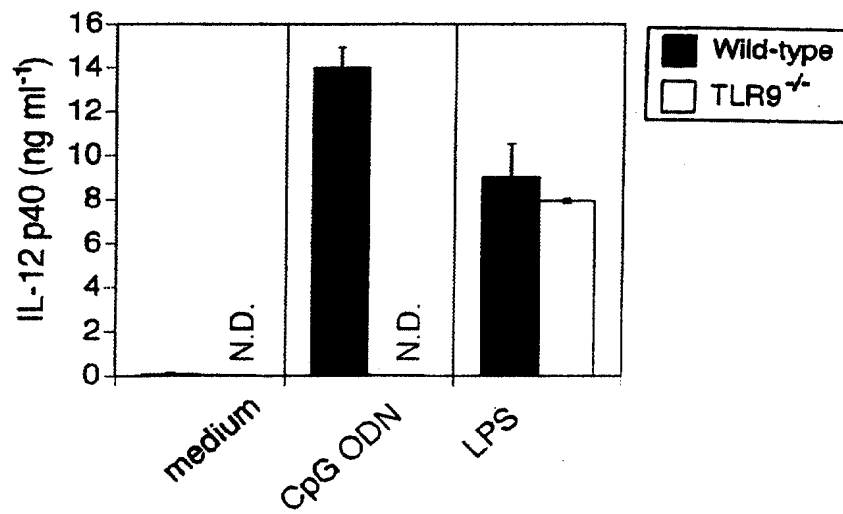


FIG. 8

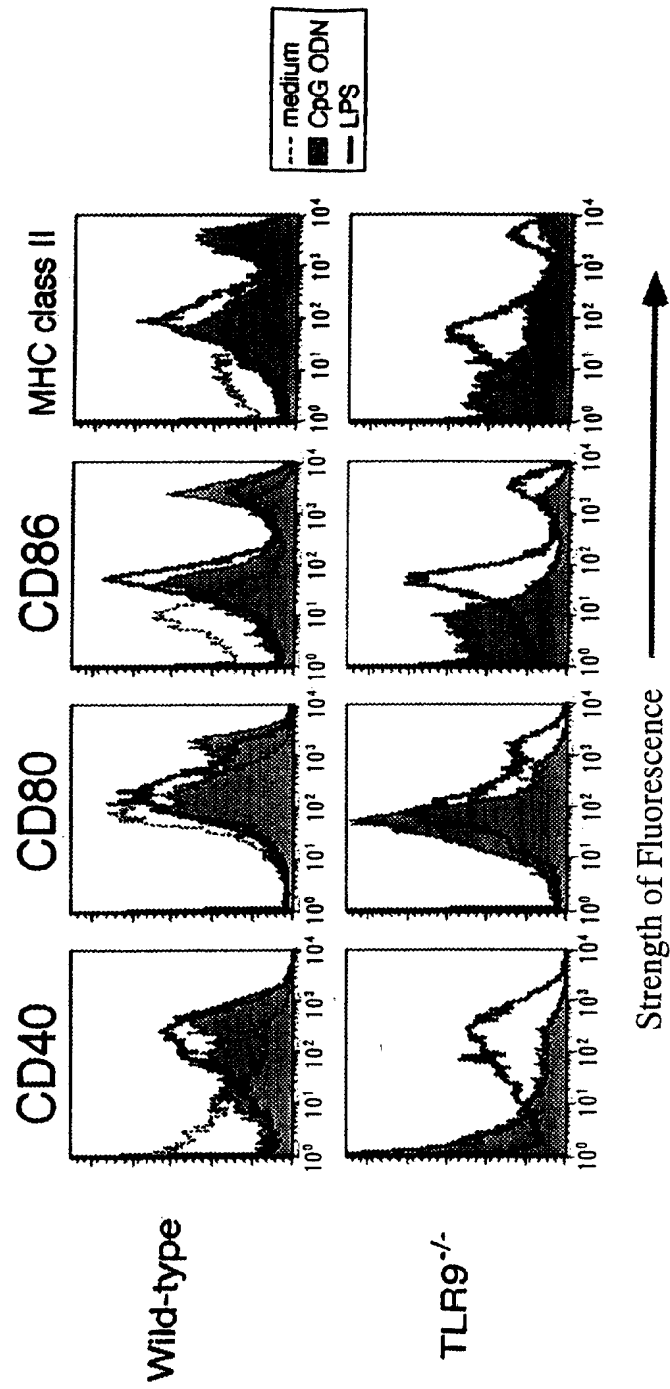


FIG. 9

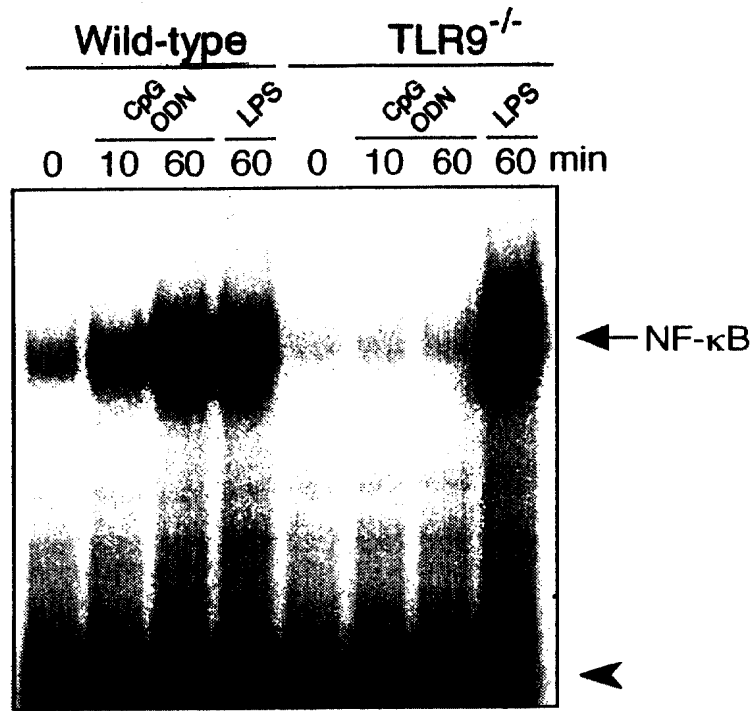


FIG. 10

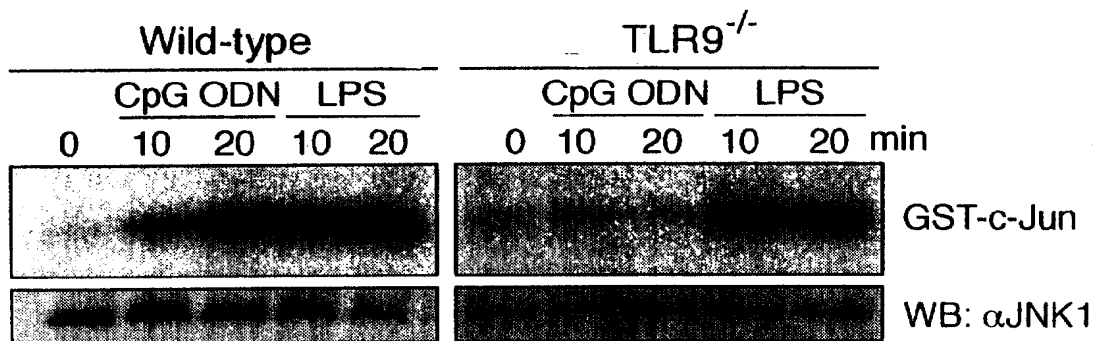


FIG. 11

